Improved Accuracy, Precision and Laboratory Productivity from Employing a Single Tube SPE-GCB Sample Preparation Configuration for the Analysis of PFAS under DOD QSM 5.3



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- BRIEF History of PFAS
- Overview of Current Methodology
- SPE requirements for EPA Methods (Drinking Water)
- Non-DW matrices DOD requirements
- Future considerations
- Conclusion



Accidental Chemicals

- Viagra
- Penicillin
- Saccharin
- Teflon





Negative impact of PFAS

Forever chemicals

- Biopersistent hard to destroy (remediation)
- Bioaccumulative/Biomagnified
- Highly mobile
- Toxic???

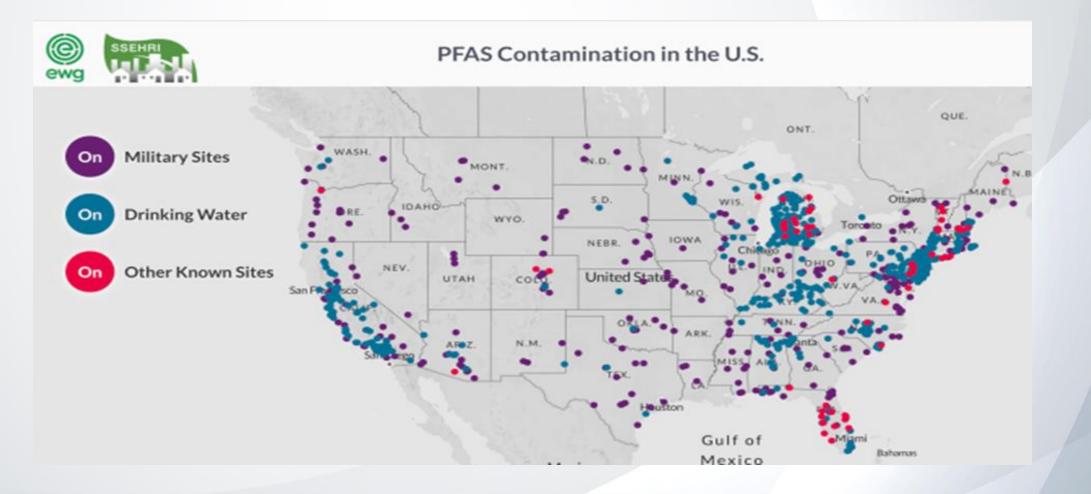


Negative Impact of PFAS

Toxicity???

- Animal studies show possible health issues
- Lack of any definitive data to link PFAS (esp. specific PFAS compounds)
- Different PFAS have different effects (see Beluga whale study)
- Short chain with shorter half life





https://www.ewg.org/interactive-maps/pfas_contamination/





https://www.pennlive.com/news/2021/02/pa-american-watersues-manufacturer-of-toxic-pfas-forever-chemicals.html



History of PFAS Analytical methodology

- EPA 537 (2009)
- EPA 8327 (2015) 2021
- EPA 537.1 (2018)
- EPA 533 (2020)
- DOD QSM (2019 for Ver. 5.3) NOT A METHOD!
- No official/promulgated methods for non-DW matrices (NOT TRUE AS OF LAST WEEK!)



History of PFAS Analytical methodology

- EPA 537 (Polymer SPE)
- EPA 537.1 (SDVB SPE only)
- ASTM D7979/EPA 8327 (DAI)
- EPA 533 (WAX SPE)
- DOD (WAX + GCB)
- No official/promulgated methods for non-DW matrices



Solid Phase Extraction Protocol

Following the procedures of EPA Method 537.1, V2, Sections 6.9 - 6.11 and 11.3 - 11.4

Cartridge: Strata SDB-L, 500 mg/6 mL Part No.: <u>8B-S014-HCH</u> Load: 250 mL sample that has been fortified with surrogates Elution: 2x 3 mL Methanol Dry Down: With Nitrogen in a heated water bath Reconstitute: Adjust final volume to 1 mL with 96:4 Water:Methanol (v/v) and add internal standards

Method Precision and Accuracy from the Analysis of 50 Co Laboratory Control Samples (LCS)

	Mean % Recovery	Standard Deviation
13C2-PFDA	106	15.8
13C2-PEHXA	108	18.2
		19.8
d5-E1FOSAA	104	
13C2-HFPO-DA	104	17.1
11CI-PF3OUdS	105	10.9
9CI+PF3OUNS	104	11.9
ADONA	103	13.6
Et-FOSAA	111	13.8
HEPO-DA	104	15.6
Me-FOSAA	113	18.5
PFBS	104	14.7
PFDA	106	12
PFDoA	105	17.4
PFHpA	111	14.8
PFHxA	109	14.1
PFHxS	108	15.1
PFNA	109	12.7
PFOA	109	12.5
PFOS	111	13
PFTeDA.	103	14.6
PFTrDA	104	13.7
PFUnA	107	13.7



Special thanks to Agustin Pierri and Weck Labs for providing this data

Sample Preparation Protocol

- Pre-treatment: 100-250 mL sample is fortified with isotopically labeled analogues కొ
 - of the method analytes
 - Cartridge: Strata-X-AW 500 mg/6 mL
 - Part No.: 8B-S038-HCH
 - Load: Pass pre-treated sample through the cartridge
 - Wash 1: Aqueous Ammonium acetate followed by Methanol
 - Wash 2: Methanol
 - Elute: Ammonium hydroxide in Methanol
 - Dry Down: Under a gentle stream of Nitrogen in a heated water bath
 - Reconstitute: Adjust the final volume to 1 mL with 20 % Water in Methanol (v/v) before analyzing by LC-MS

Table 6.

EPA 533 Precision and Accuracy Data from a Commercial Laboratory

Analyte	MS	MSD	BS	BSD
11CI-PF30UdS	85%	84%	95%	86%
4-2FTS	113%	104%	109%	100%
6-2 FTS	94%	96%	108 %	102%
8-2 FTS	97 %	100%	89%	101%
9CI-PF30UdS	101 %	107%	99%	119%
ADONA	118%	116%	111%	99%
HFPO-DA	100 %	97%	110%	101 %
NFDHA	117%	126%	117%	114%
PFBA	102 %	116%	89%	95%
PFBS	117%	106%	97%	105%
PFDA	102%	99%	112%	104%
PFDoA	104%	107%	108%	109%
PFEESA	116%	109%	119%	115%
PFHpA	112%	115%	94%	97%
PFHpS	119%	117%	119%	114%
PFHxA	113%	107%	91 %	95%
PFHxS	96%	101 %	108%	110%
PFMBA	106%	101 %	111%	118%
PFMPA	99%	100 %	108%	117%
PFNA	107%	104%	105%	110%
PFOA	101%	104%	101%	100%
PFOS	117%	115%	108%	108%
PFPeA	97%	96%	92 %	88%
PFPeS	86%	99%	103%	104%
PFUnA	105%	103%	115%	113%

Special thanks to Agustin Pierri and Weck Labs for providing this data



DoD/DOE QSM 5.3 Appendix B 2019

QC Check	Minimum Frequency	Acceptance Criteria	Corrective Action	Flagging Criteria	Comments
AFFF and AFFF Mixture Samples Preparation	Each sample and associated batch QC samples.	Each field sample must be prepared in duplicate (equivalent to matrix duplicate). Serial dilutions must be performed to achieve the lowest LOQ possible for each analyte.	NA.	NA.	Adsorption onto bottle is negligible compared to sample concentration so subsampling is allowed. Multiple dilutions will most likely have to be reported in order to achieve the lowest LOQ possible for each analyte.
Sample Cleanup Procedure	Each sample and associated batch QC samples. Not applicable to AFFF and AFFF Mixture Samples.	ENVI-Carb TM or equivalent must be used on each sample and batch QC sample.	NA	Flagging is not appropriate.	Cleanup should reduce bias from matrix interferences.



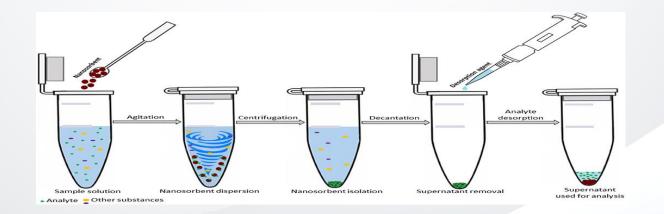
GCB in PFAS sample prep

- Only for non-DW matrices
- Removes Organic Acids that can suppress ionization (TDCA and PFOS)
- Table B-15 provides for equivalent to Envi CarbTM
- Can actually improve recovery of long chain/neutral compounds (depends on format)
- Potential loss of analytes unless GCB is re-extracted
- Several ways to skin a cat!



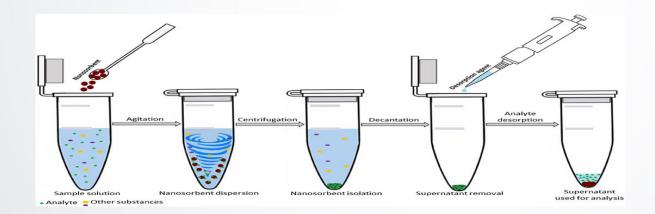
GCB for QSM 5.3, Table B-15

dSPE with Loose GCB (10mg-100mg)





Downside of dSPE



- Time additional 45-60 minutes per batch of samples
- Precision more sample manipulation = more variability
- Messy Loose GCB requires use of PPE and ventilation
- Recovery potential loss of long chain compounds (unless re-extract the GCB)
- Potential clogging of LC or MS from contamination with GCB from extract



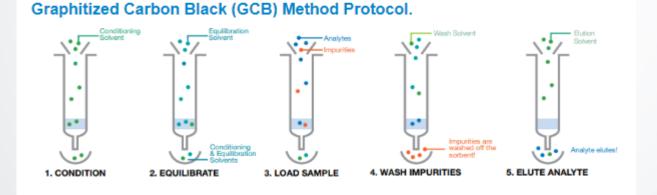
Analyte	WAX SPE +dSPE GCB % Recovery	Strata PFAS Stacked Cartridge % Recovery
13C2-PFDoDA	77	84.5
13C2-PFTeDA	62	84.0
PFODA	38	78.3
PFHxDA	63	89.3

Table 1: Recovery Comparisons of WAX SPE and dSPE using GCB vs Strata PFAS Single Cartridge Method

Special thanks to Eurofins Lancaster Labs for providing this data



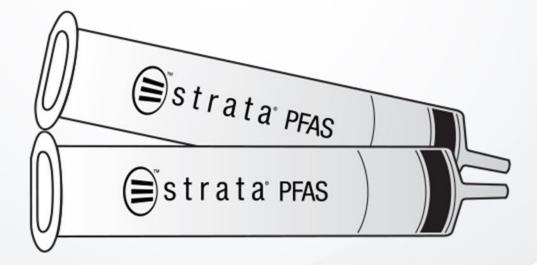
Downside of separate GCB tubes



- Additional Time approximately 15 minutes per batch of samples
- Cost solvent and tube (\$5-7 per sample)



Incorporate WAX and GCB in a single SPE tube!



- 200mg WAX/50mg GCB
- 500mg WAX/50mg GCB



		dSPE		Sta	cked Tubes CS0-9207	
Analyte	Spk Conc. (ng/L)	% Rec	RSD	Spk Conc. (ng/L)	% Rec	RSD
10:2 FTS	50.00	80%	6.2%	20.00	94%	11.5%
11CI-PF3OUdS	50.00	85%	23.6%	20.00	93%	7.0%
3:3 FTCA	50.00	89%	11.6%	20.00	86%	4.2%
4:2 FTS	50.00	100%	2.9%	20.00	103%	2.9%
5:3 FTCA	50.00	86%	0.6%	20.00	94%	3.0%
6:2 FTS	50.00	98%	5.0%	20.00	109%	4.5%
7:3 FTCA	50.00	79%	12.6%	20.00	90%	5.3%
8:2 FTS	50.00	97%	5.8%	20.00	105%	3.4%
9CI-PF3ONS	50.00	94%	18.6%	20.00	95%	5.9%
ADONA	50.00	99%	3.2%	20.00	100%	3.4%
EtFOSA	50.00	109%	9.6%	20.00	104%	11.3%
EtFOSE	50.00	92%	11.8%	20.00	92%	7.1%
HFPO-DA	50.00	110%	6.9%	20.00	102%	9.9%
MeFOSA	50.00	108%	6.6%	20.00	102%	16.7%
MeFOSE	50.00	93%	11.2%	20.00	109%	8.4%
N-EtFOSAA	50.00	96%	6.1%	20.00	101%	11.2%
N-MeFOSAA	50.00	103%	9.7%	20.00	99%	12.1%
PFBA	50.00	96%	1.4%	20.00	96%	0.6%
PFBS	50.00	97%	3.2%	20.00	98%	4.7%
PFDA	50.00	101%	4.3%	20.00	97%	6.1%
PFDoDA	50.00	100%	1.7%	20.00	98%	3.6%
PFDS	50.00	85%	21.5%	20.00	96%	6.9%
PFHpA	50.00	99%	2.8%	20.00	97%	3.2%
PFHpS	50.00	102%	1.9%	20.00	92%	6.1%
PFHxA	50.00	96%	2.3%	20.00	100%	5.4%
PFHxDA	50.00	73%	15.6%	20.00	97%	1.0%
PFHxS	50.00	97%	0.8%	20.00	95%	7.3%
PFNS	50.00	97%	10.5%	20.00	95%	3.7%
PFOA	50.00	106%	8.0%	20.00	101%	3.8%
PFOcDA	50.00	32%	23.8%	20.00	87%	2.5%
PFOS	50.00	96%	12.5%	20.00	98%	5.0%
PFPeA	50.00	96%	3.6%	20.00	98%	4.0%
PFPeS	50.00	95%	4.2%	20.00	95%	5.7%
PFTeDA	50.00	100%	2.6%	20.00	100%	4.2%
PFTrDA	50.00	96%	12.5%	20.00	94%	2.2%
PFUndA	50.00	104%	5.9%	20.00	97%	0.8%
Average		94%	8%		98%	6%



Analyte	Actual	Sample Result	% Recovery	Method Recommendation Limits	Pass/Fail
	Concentration				
PFBA	25.600	22.640	88	84-135	Pass
PEPeA	25.600	22.157	87	75-138	Pass
PFBS	22.640	22.300	99	81-133	Pass
4:2-FTS	23.920	22.078	92	64-134	Pass
PFHsA	25.600	24.644	96	80-137	Pass
PFPeS	24.000	21.699	90	82-132	Pass
HEPODA	25.600	26.336	103	70-130	Pass
PFHpA	25.600		106	80-140	Pass
PFHxS	24.200	913	102	71-131	Pass
DONA	24.120	26.083	108	70-130	Pass
6:2-FTS	24,280	24.217	100	51-155	Pass
PFHpS	24.360	23.015	94	80-129	Pass
PFOA	25.600	25.043	98	83-138	Pass
PEQS	24,480	22,492	92	54-139	Pass
PENA	25.600	25.872	101	73-140	Pass
9CI-PF3ONS	23.840	21.863	92	70-130	Pass
PENS	24.560	21.993	90	71-121	Pass
PFDA	25.600	25.047	98	78-137	Pass
8:2-FTS	24.520	22.231	91	62-133	Pass
PFOSA	25.600	25.714	100	73-121	Pass
NMEFOSAA	25.600	30.906	121	53-136	Pass
PFDS	24.640	22.873	93	69-124	Pass
PFUnDA	25.600	26.353	103	70-134	Pass
NEUFOSAA	25.600	28.765	112	59-145	Pass
11CI-PF3OUdS	24.120	22.625	94	70-130	Pass
PFDeDA.	25.600	27.710	108	75-139	Pass
10:2-FTS	24,680	26.626	108	50-124	Pass
PFDo5	24.800	21.509	87	39-121	Pass
PETrDA	25.600	25.814	101	67-144	Pass
PFTeDA	25.600	25.446	99	79-134	Pass
PEHADA	25.600	29.662	116	36-136	Pass
PFODA	25.600	27.373	107	10-124	Pass

Table 2: Recovery of QSM 5.3 Target Analytes from a Laboratory Control Sample Using Strata PFAS SPE (WAX/GCB)

Recovery Range: 87 % - 116 % Average Recovery: 98.8 % Mean Recovery: 99.0 %



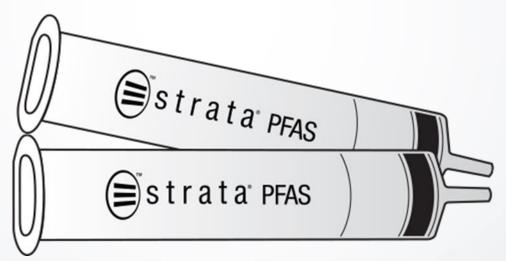
Special thanks to Eurofins Lancaster Labs for providing this data

Percent Recoveries of a 2ng/L for Matrix Spikes (MS) and Matrix Spike Duplicates (MSD) of PFAS compounds spikes in a wastewater Discharge pond, Stormwater Runoff and Two Wastewater effluents.

	ww.r	Effluent 1	WW Disc	harge Pond	Stormwat	er Runoff	WW Fff	uent 2
	MS	MSD	MS	MSD	MS	MSD	MS	MSD
	94%	92%	86%	92%	109%	114%	118%	123%
	5478	52,0	00%	5278	10576	227/0	110/0	11.370
	112%	115%	100%	117%	112%	115%	120%	136%
	209%	219%	239%	218%	162%	182%	2040%	2003%
							111%	106%
	104%	79%	123%	123%	103%	113%	108%	114%
	107%	70%	133%	139%	106%	119%	114%	115%
	125%	76%	166%	163%	102%	103%	111%	119%
	107%	82%	106%	107%	93%	114%	122%	112%
	103%	96%	145%	160%	117%	88%	112%	101%
	96%	71%	121%	114%	93%	113%	126%	117%
	99%	94%	104%	87%	102%	96%	105%	110%
	109%	94%	102%	108%	96%	102%	109%	114%
	113%	85%	104%	110%	99%	95%	117%	125%
	110%	90%	97%	95%	109%	114%	113%	114%
	117%	92%	83%	88%	114%	110%	119%	123%
	105%	88%	110%	114%	129%	133%	102%	106%
	104%	83%	71%	74%	89%	88%	95%	96%
	93%	86%	78%	92%	82%	86%	94%	97%
	107%	94%	67%	67%	116%	108%	118%	127%
	104%	110%	54%	56%	100%	103%	116%	125%
	90%	101%	54%	55%	65%	63%	92%	105%
	114%	76%	126%	134%	91%	93%	107%	124%
	122%	77%	131%	144%	102%	98%	107%	106%
	109%	80%	104%	99%	87%	101%	115%	86%
	105%	93%	90%	106%	85%	86%	92%	95%
	83%	77%	92%	94%	107%	100%	103%	102%
	63%	1170	9276	9476	10776	100%	103%	10276
	70%	66%	55%	51%	92%	90%	95%	91%
	102%	100%	92%	72%	120%	102%	142%	161%
	86%	106%	64%	83%	124%	102%	130%	118%
	96%	100%	54%	61%	112%	105%	113%	124%
	100%	97%	67%	72%	110%	112%	137%	146%
	127%	95%	116%	101%	120%	110%	120%	114%
	117%	84%	109%	118%	98%	102%	112%	109%
	106%	91%	68%	115%	101%	98%	102%	111%
	82%	86%	51%	69%	95%	83%	96%	96%
	119%	93%	109%	111%	107%	104%	114%	111%
	114%	97%	106%	134%	129%	104%	128%	107%
	107%	105%	70%	69%	47%	54%	80%	87%
	101%	94%	84%	83%	83%	71%	23%	54%
	108%	120%	97%	95%	98%	112%	111%	106%
	121%	126%	102%	116%	132%	129%	134%	133%
	117%	103%	107%	116%	114%	112%	125%	123%
	117%	103%	96%	108%	114%	103%	125%	
					104/8	10376	11/70	120%
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Special thanks to David Scheissel and Babcock Labs for providing this data

BENEFITS OF STACKED TUBES



- High recoveries that meet DOD requirements
- No need for PPE (N95 Masks or hoods)
- Lower cost of solvents and consumables
- Less sample manipulation
- Consistent formats for reduced variability
- Single step extraction and cleanup to save time and reduce costs



Future considerations

- UCMR5
- GCB Pass Through (50mg or 25mg)
- On-line SPE (Including On-line GCB cartridges)
- Unique SPE formats for dirty matrices (soil, sediment, biota)
- Extended panels and resolution issues Unique LC chemistry and MP needed to improve resolution of more analytes
- TOP and TOF assays for fingerprinting source and showing transport mechanisms
- Non-targeted assays
- Pending 1600 Series method to replace Table B15



Conclusion

- 1. PFAS are not going away and we have just scratched the surface
- 2. Lots of different ways to analyze PFAS depending on matrix and analyte list
- 3. Stacked Tubes offer performance and productivity benefits for DOD compliant work
- 4. Phenomenex SPE products are demonstrated Fit for Purpose for all published methods
- 5. New formats and chemistries being considered
- 6. https://www.phenomenex.com/Info/Page/pfasresources



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Questions?

