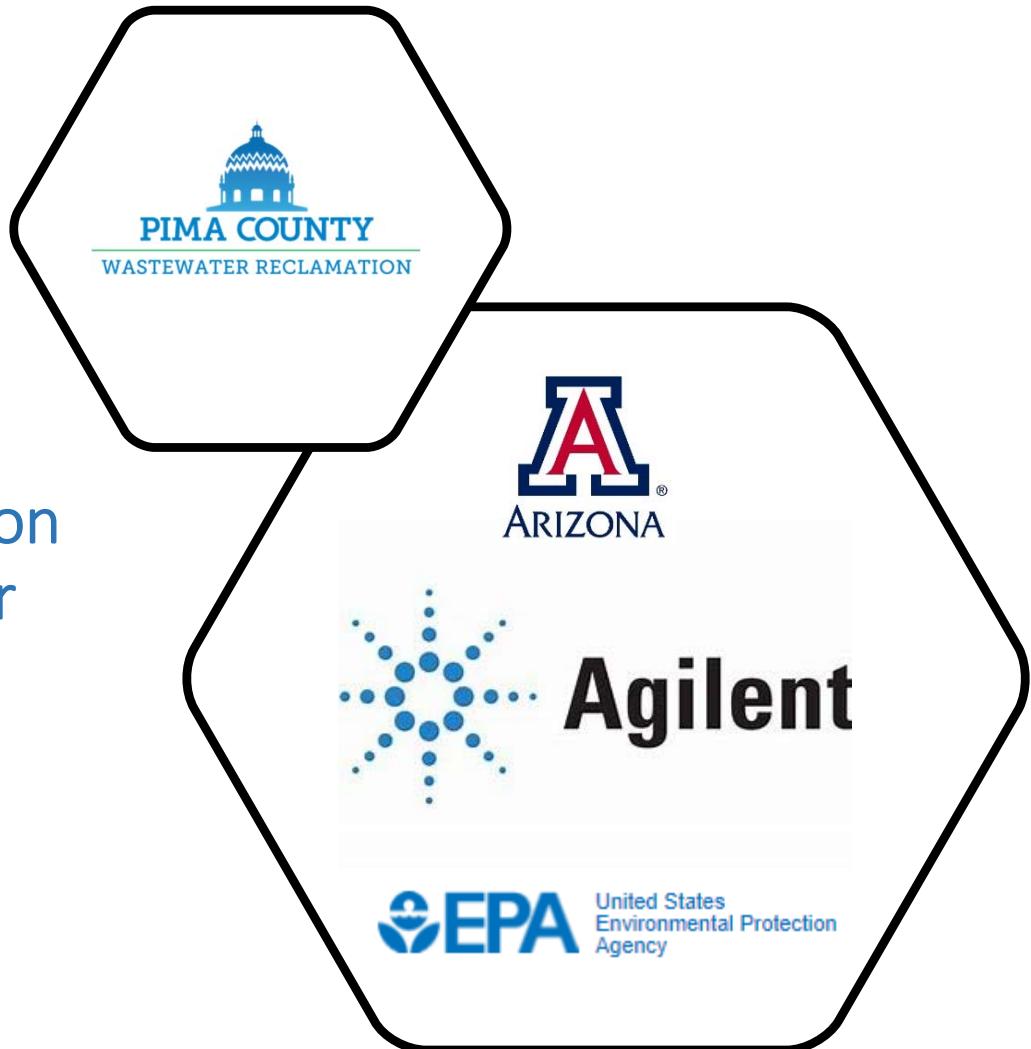


# Standardized Method for the Identification and Quantification of Microplastics in Wastewater Using LDIR Microscopy

By

Jeff Prevatt



# Recent Headlines

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PLASTICS

## Microplastics Detected in Human Stool Samples for First Time

By Lorraine Chow | Oct. 23, 2018 09:07AM EST

Powered by RebelMouse

HEALTH + WELLNESS



Microplastics. [MPCA Photos / Flickr / CC BY-NC 2.0](#)

OCTOBER 31, 2018

## Is there a risk to human health from microplastics?

by BfR Federal Institute for Risk Assessment

According to EFSA, there is a possibility of oral absorption of microplastic particles of a certain size, although the fate and possible degradation in the *gastrointestinal tract* have not been sufficiently investigated so far due to a lack of analytical methods and valid studies.

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## Recent Headlines

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# MICROPLASTIC **MAYHEM**

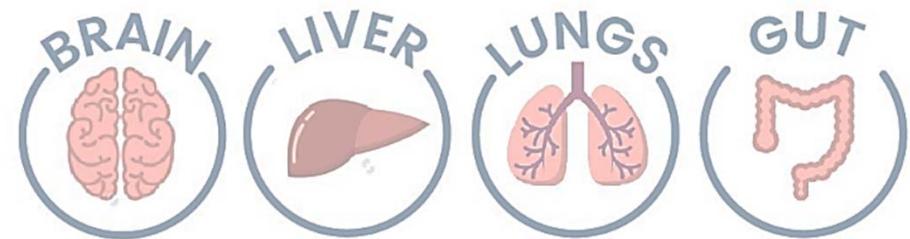
An Emerging Health Crisis

93% Percent of Bottled Water Samples Contain Microplastics

- Polypropylene
- Nylon
- Polyethylene Terephthalate

## TOP MICROPLASTIC PERILS

Toxic to:



Humans Ingest MICROPLASTICS via

- SEAFOOD • CONTAMINATED WATER
- DRINKING FROM SINGLE-USE PLASTIC BOTTLES

# Oh, yuck! You're eating about a credit card's worth of plastic every week

Doyle Rice, USA TODAY

Published 6:24 p.m. ET June 12, 2019 | Updated 1:11 a.m. ET June 13, 2019



The analysis *No Plastic in Nature: Assessing Plastic Ingestion from Nature to People* prepared by Dalberg, based on a study commissioned by WWF and carried out by University of Newcastle, Australia, suggests people are consuming about 2000 tiny pieces of plastic every week. That's approximately 21 grams a month, just over 250 grams a year.



# Standardization is Needed

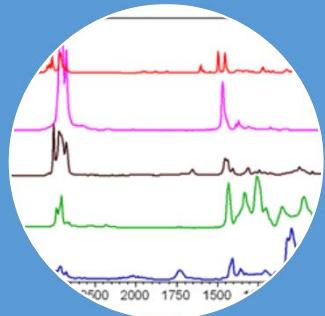
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Sample  
Collection



Sample  
Preparation



Sample  
Analysis



Reproducibility



Sample  
Collection

## Desired Features

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- Representative of source
- Account for diurnal patterns
- Volume proportional

# Wastewater Sampling Apparatus

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- 1) Pump capable of providing 20' of head and up to 3 gpm flow
- 2) Stainless steel transfer tubing
- 3) Adjustable valve for flow control
- 4) Programmable timer
- 5) Sieve Stack (1,000 µm – 20 µm)
- 6) Container for sieves with drainage

# Sampling Options

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Hose Bib Valve



Flow Meter

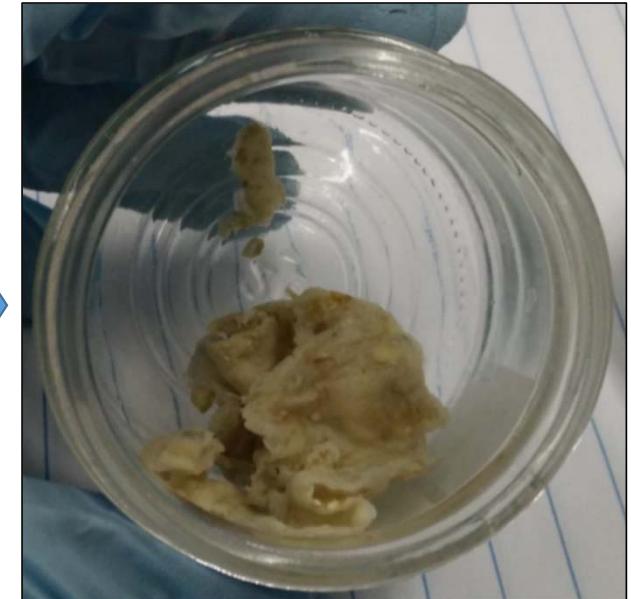


- Easy Setup & Calibration
- Improved Flow Control
- Recorded Total Volume
- Submersible or Peristaltic Pump

# Sample Retrieval

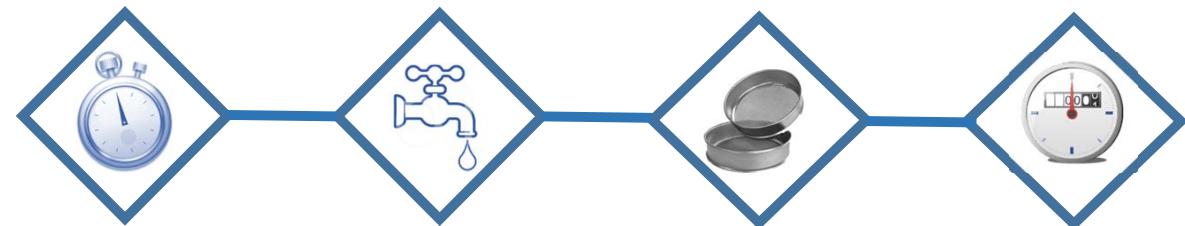
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Sieved material rinsed and collected in sample container





## Sample Collection



- Straightforward
- Simple to follow
- Reproducible



# Desired Outcome

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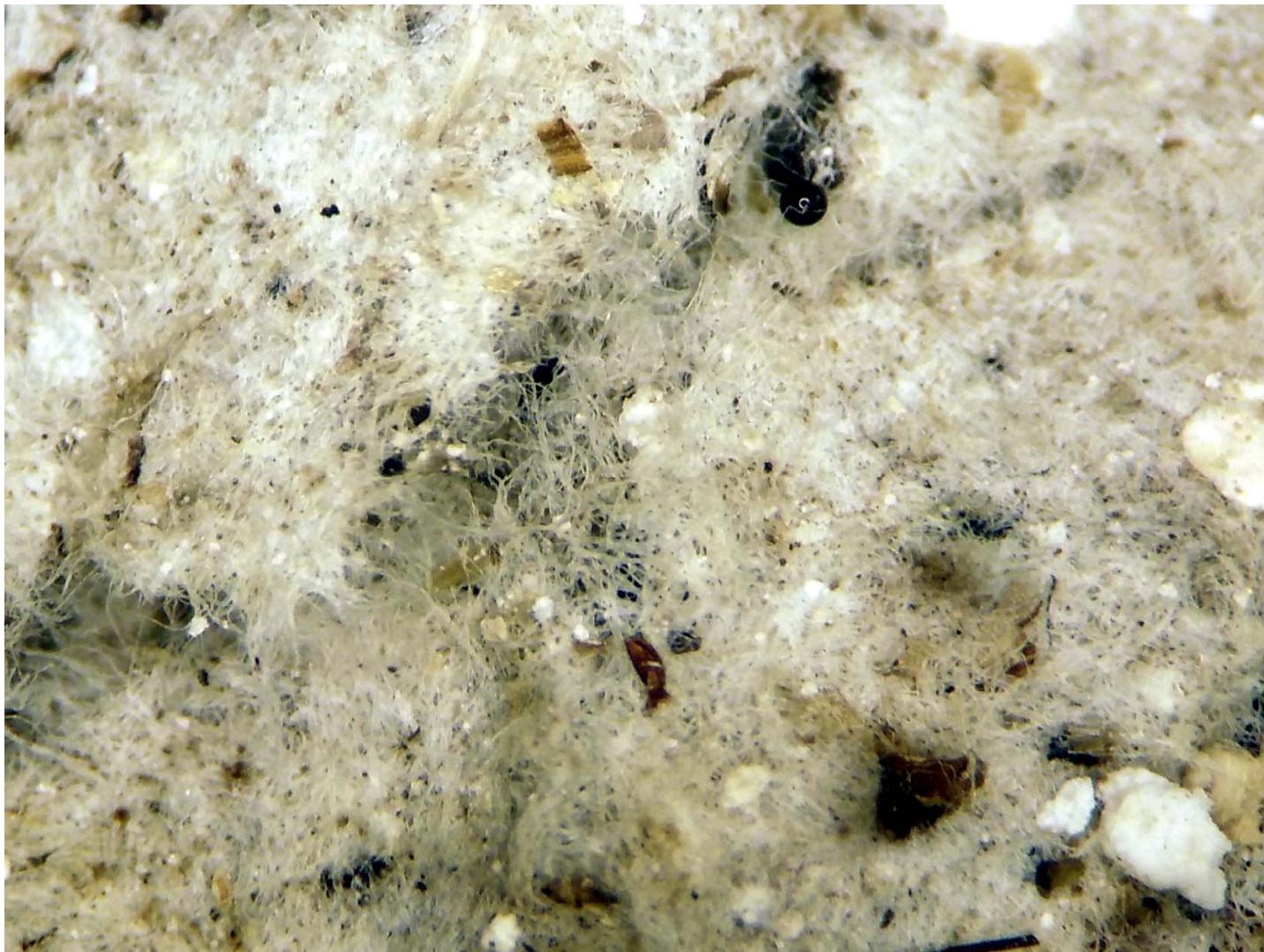


Sample  
Preparation

Procedure MUST:

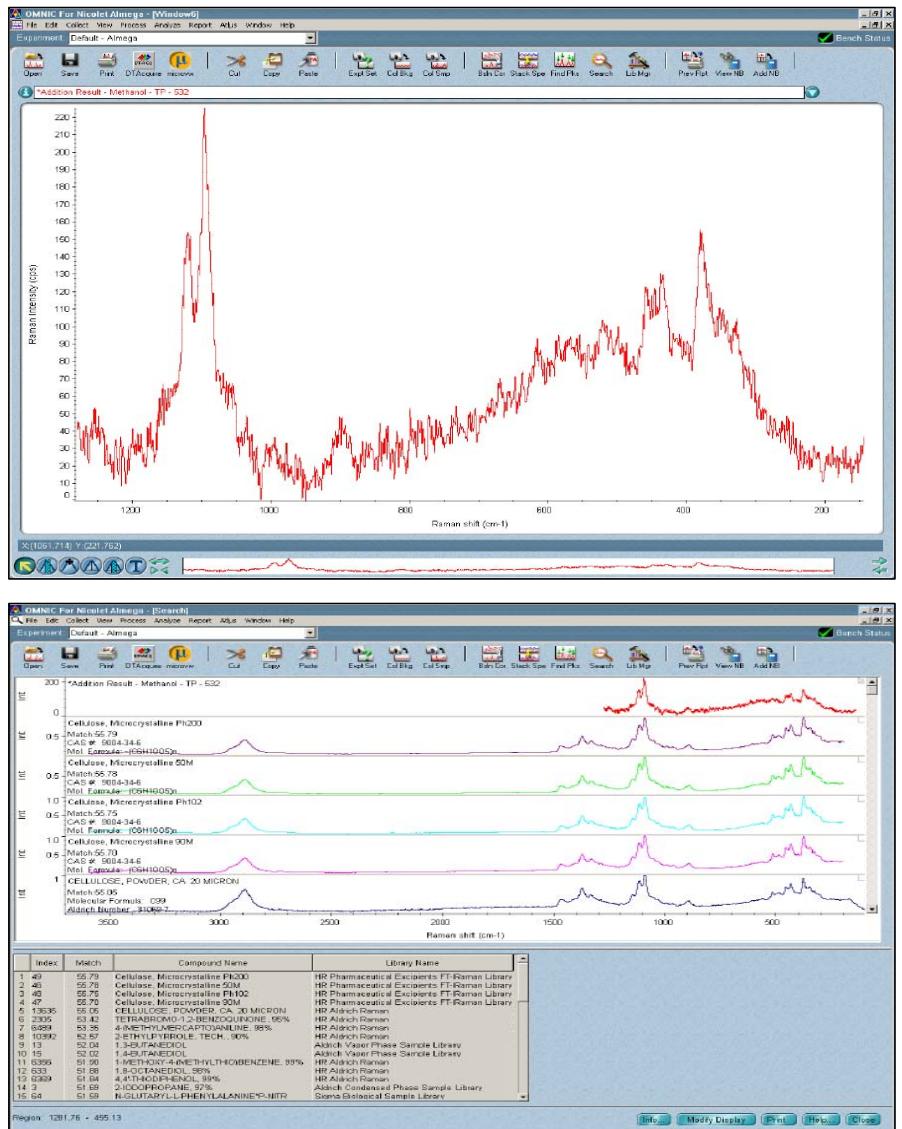
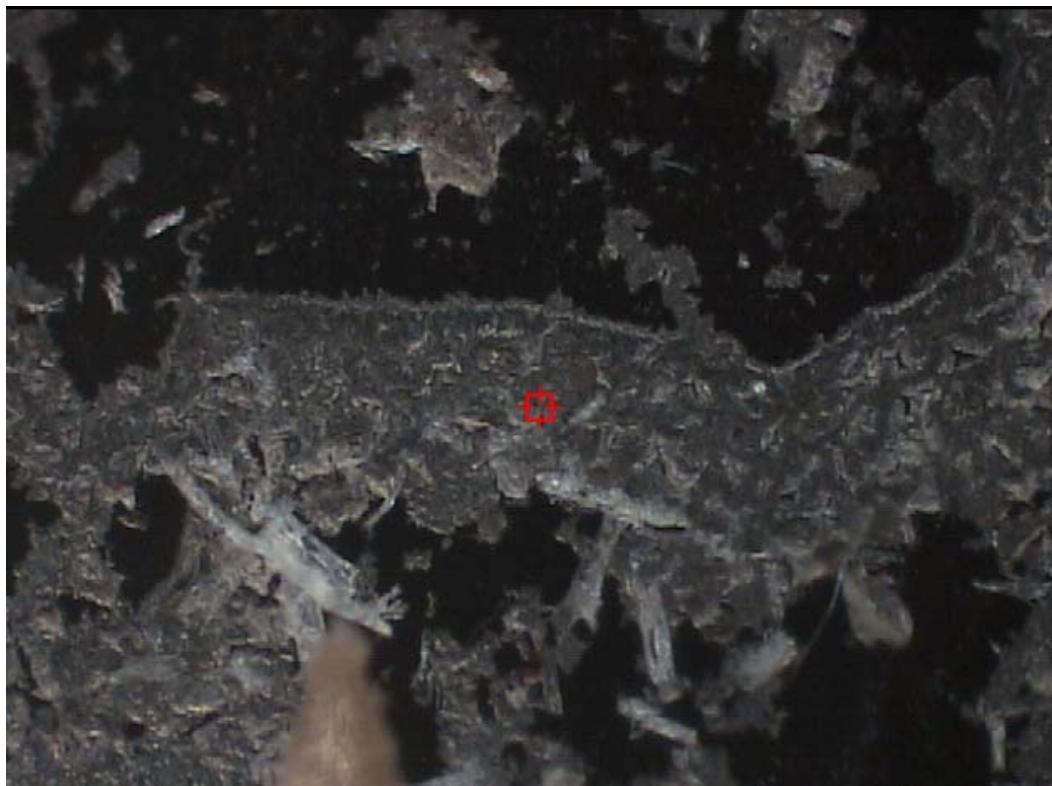
- Account for interferences
- Easy to follow & replicate
- Produce accurate results
- Minimize loss or damage of particles

~20x magnification



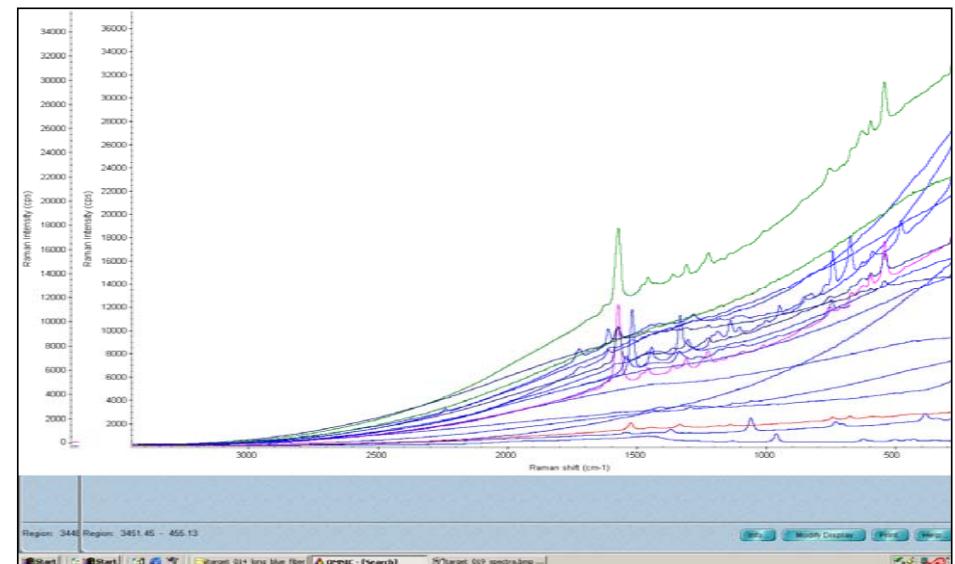
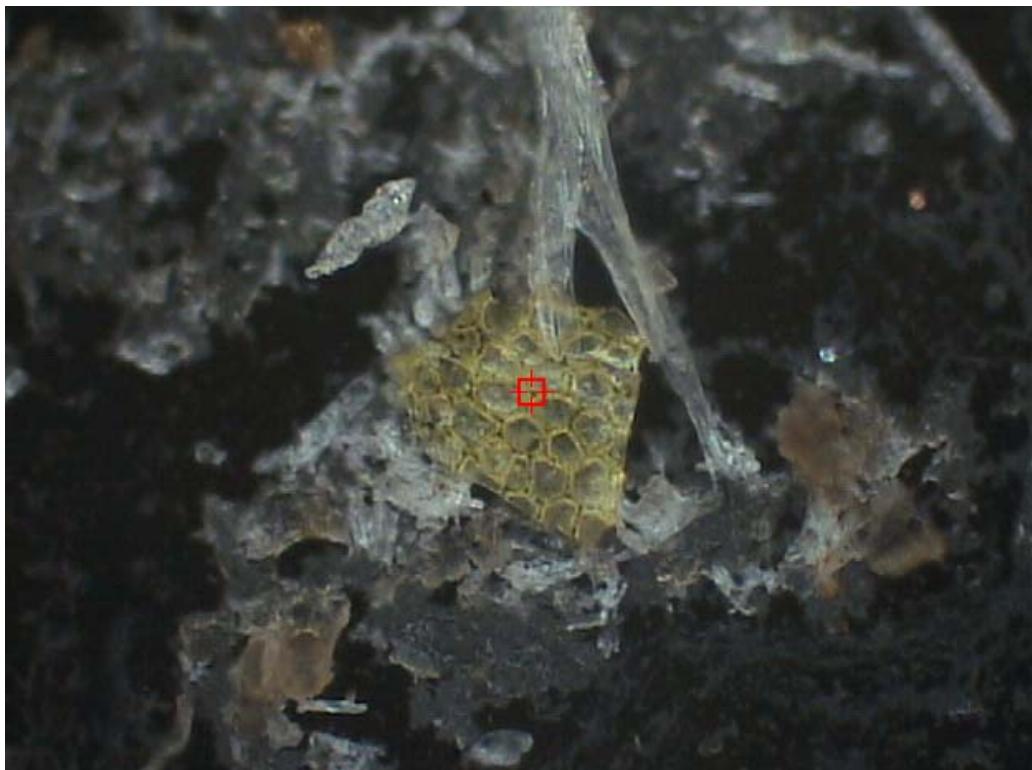
# Interferences

Cellulose is abundant  
(toilet paper)



# Interferences

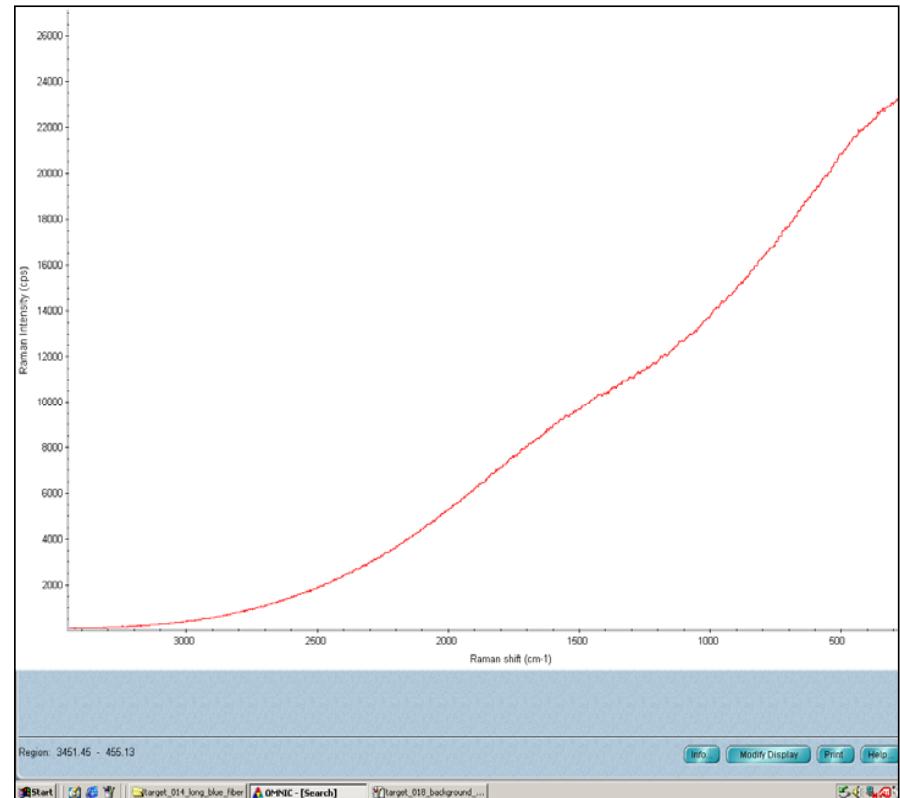
Abundant chitin background material



# Interferences

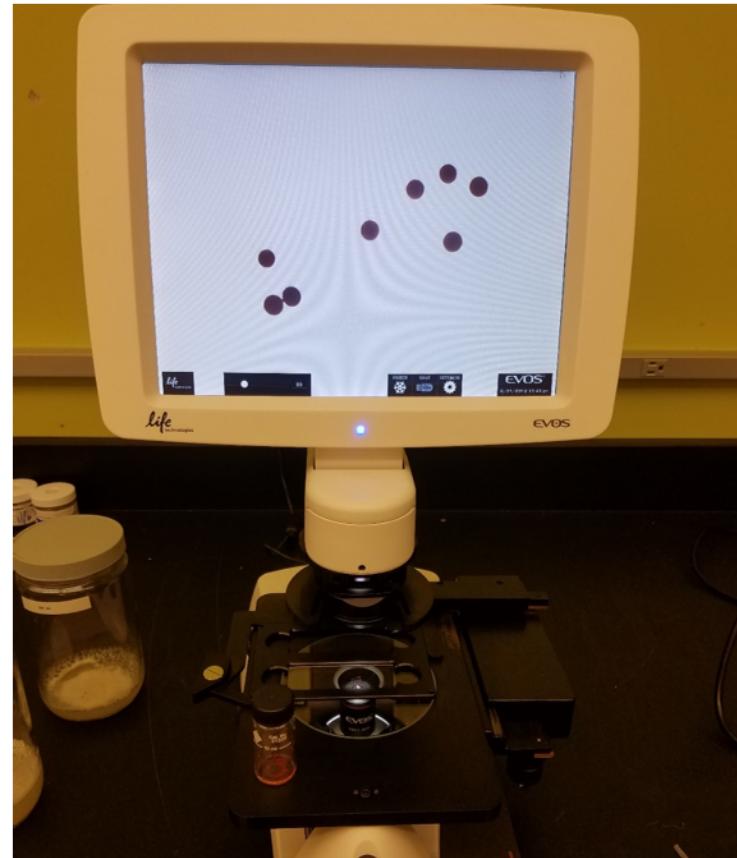
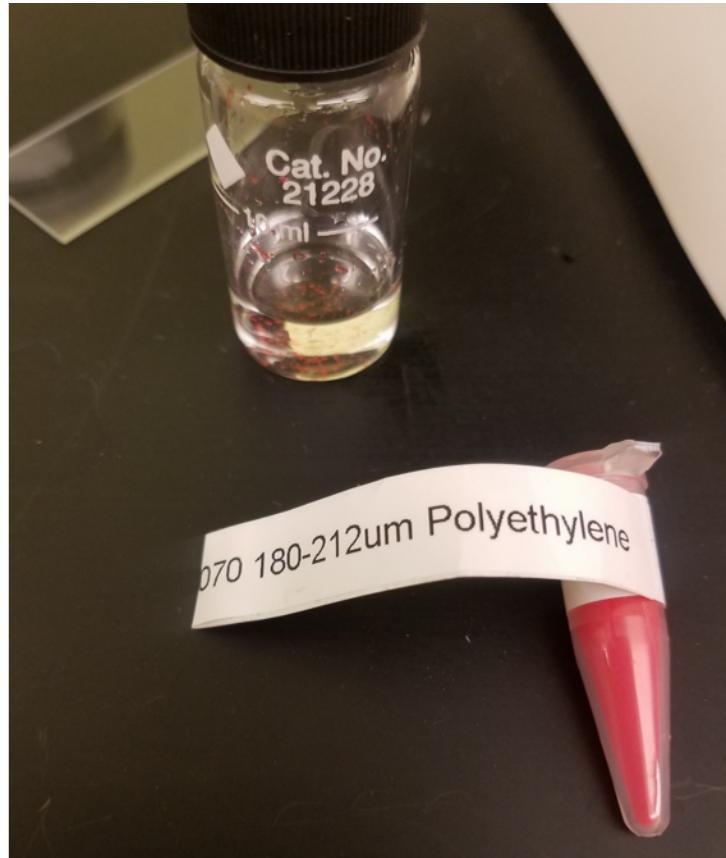
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Skin and lipid material is abundant



# Extraction Procedure Reference Spikes

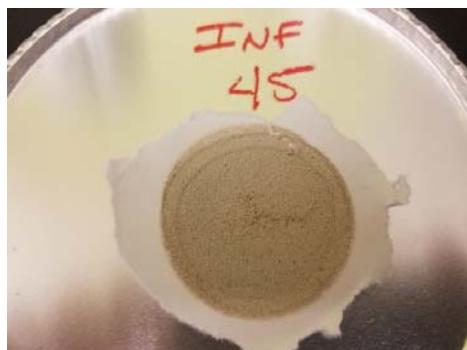
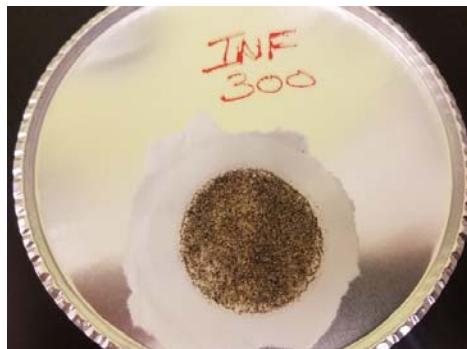
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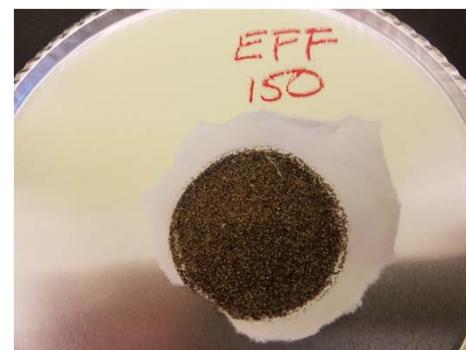
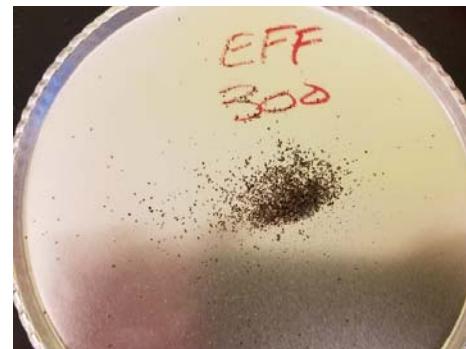
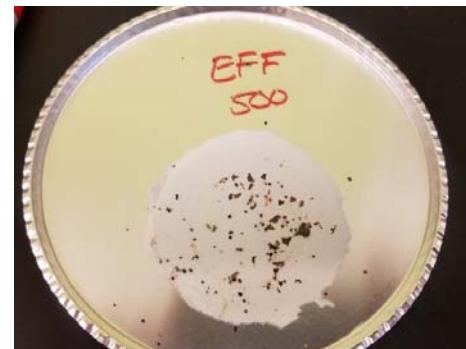
# Sieve Fractions

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Influent



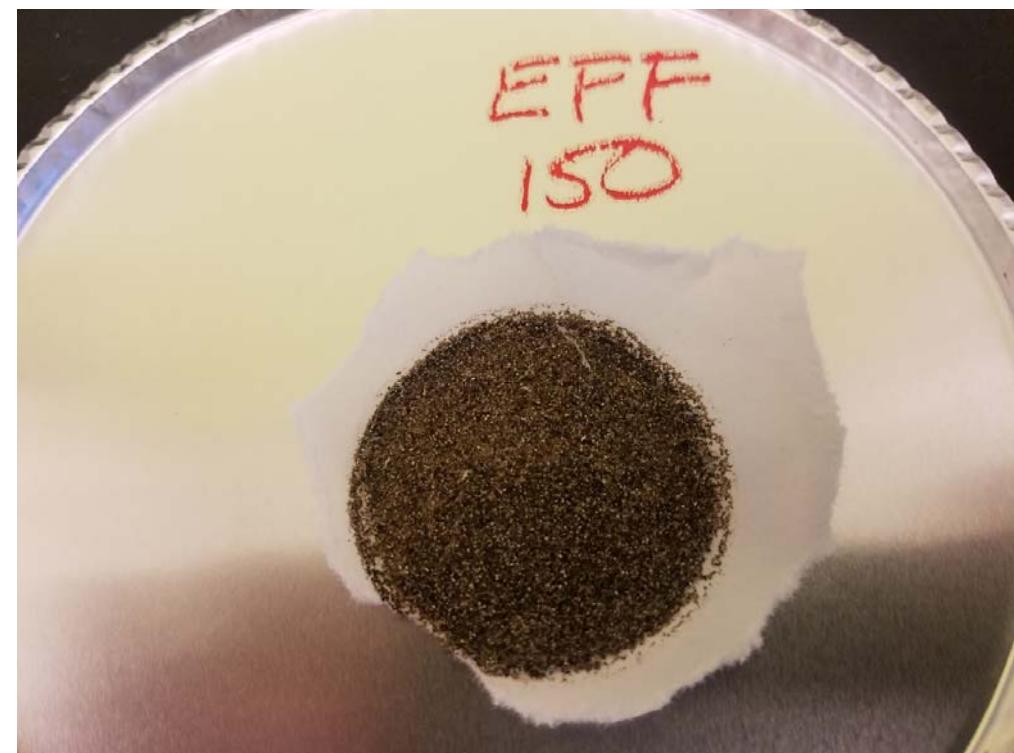
Effluent



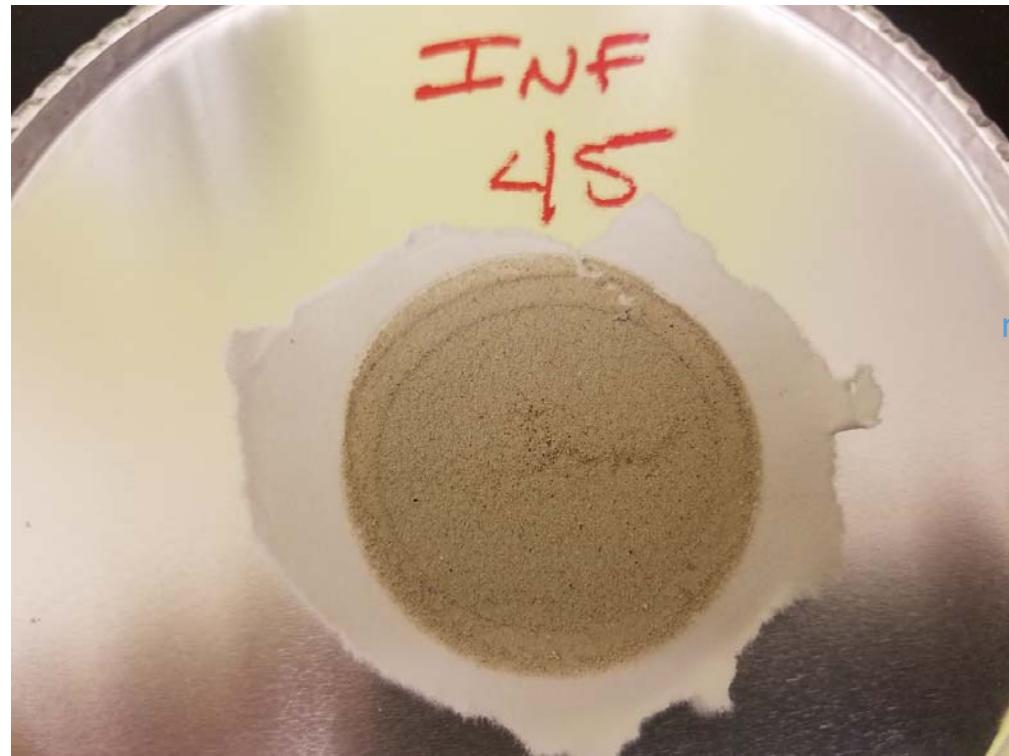
Influent 150 micron



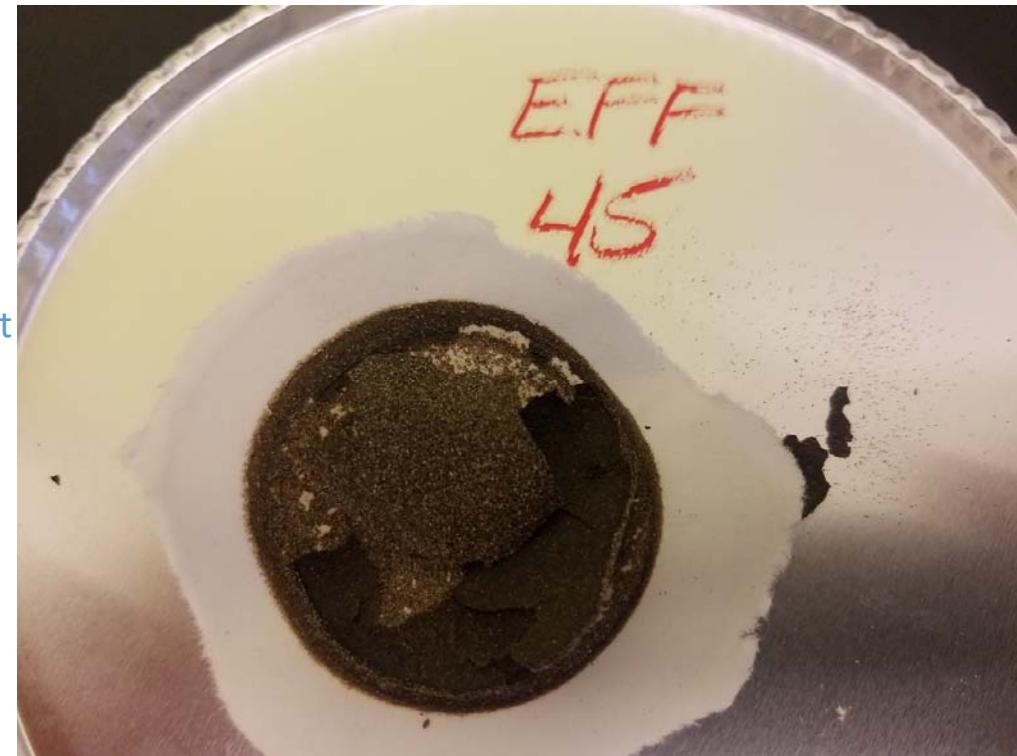
Effluent 150 micron



Influent 45 micron



Effluent 45 micron



# Universal Enzymatic Purification Protocol

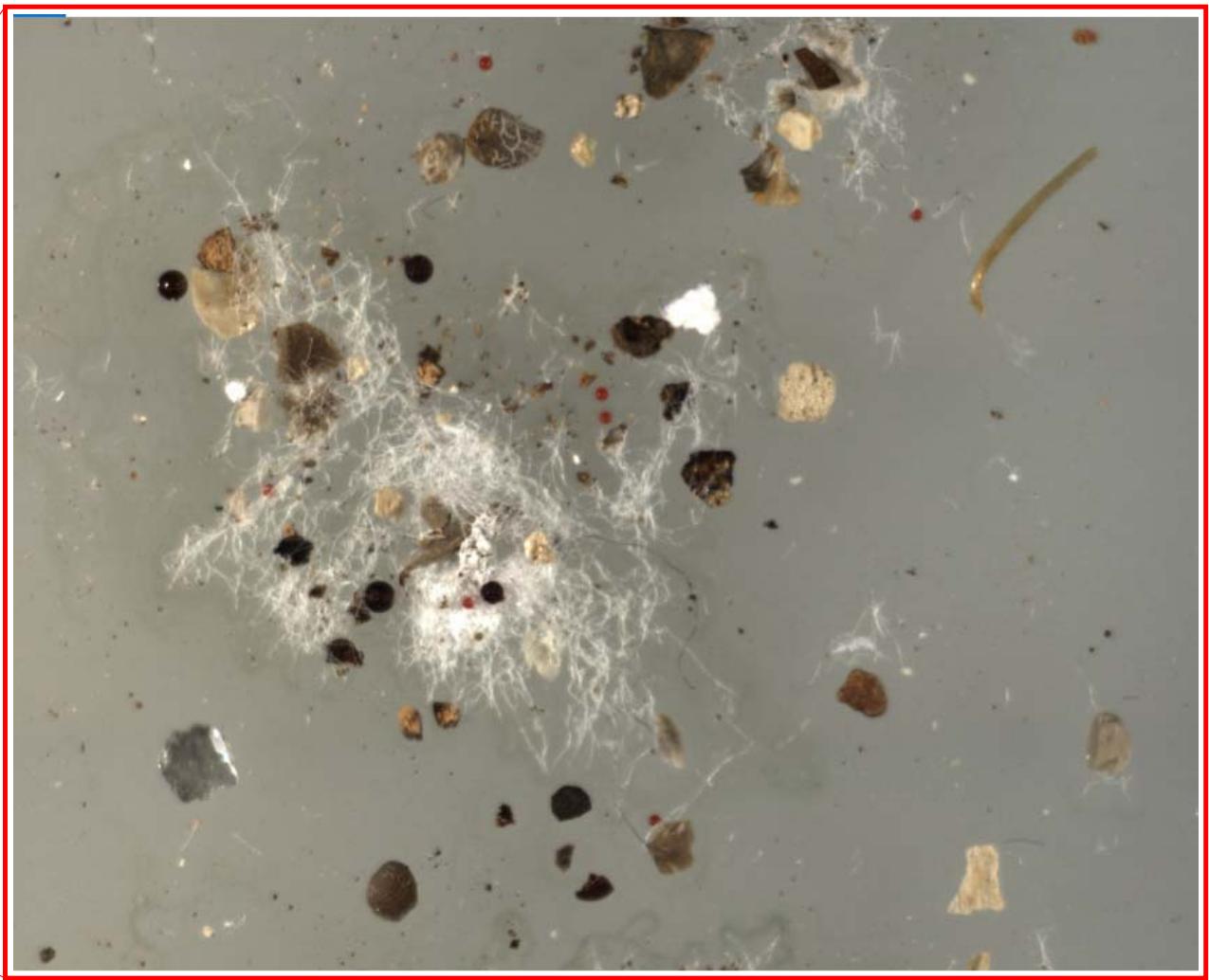
Löder 2017

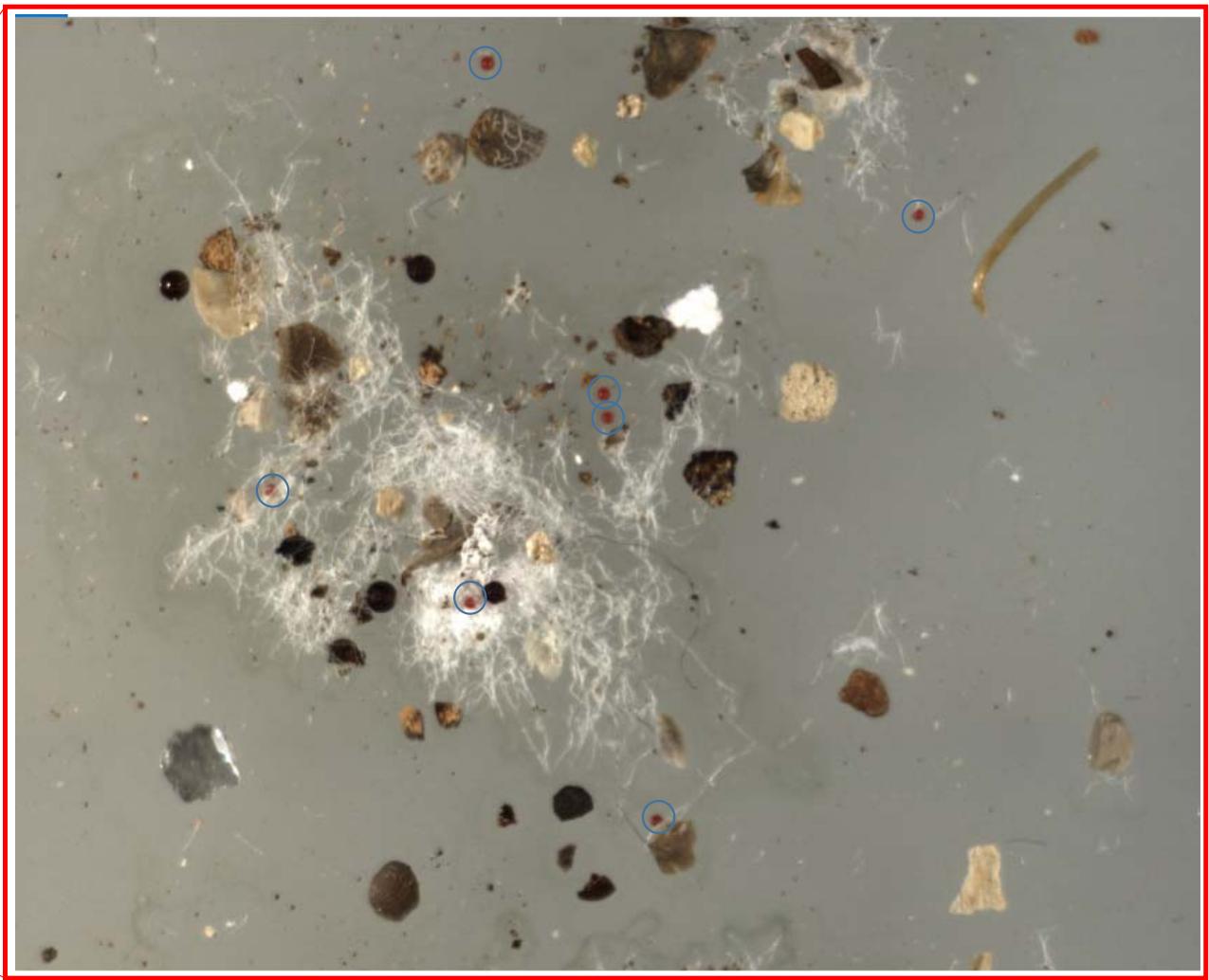
100 mL of SDS 10% (1,2,3,4,5)	1 d	50 °C	pH 6	40 °C	50 °C	37 °C
20 mL of protease (1,2,3,4) + 100 mL Tris HCl 1 M buffer	1 d					
5 mL of lipase (4) + 100 mL Tris HCl 1 M buffer	1 d					
25 mL of cellulase (1,2,3,5) + 100 mL NaOAc 1 M buffer	3 x 1 d					
20 mL of amylase (5) + 100 mL NaOAc 1 M buffer	1 d					
30 mL of H <sub>2</sub> O <sub>2</sub> I* <sup>2</sup> (1,2,3,4,5) 35%	1 d					
5 mL of chitinase (1,2,3,4) + 100 mL NaOAc 1 M buffer	3 d		pH 5			
30 mL of H <sub>2</sub> O <sub>2</sub> II* <sup>2</sup> (1,2,3,4,5) 35%	1 d					
Density separation with ZnCl <sub>2</sub> (1,2,3,4,5)	1 - 3 d					

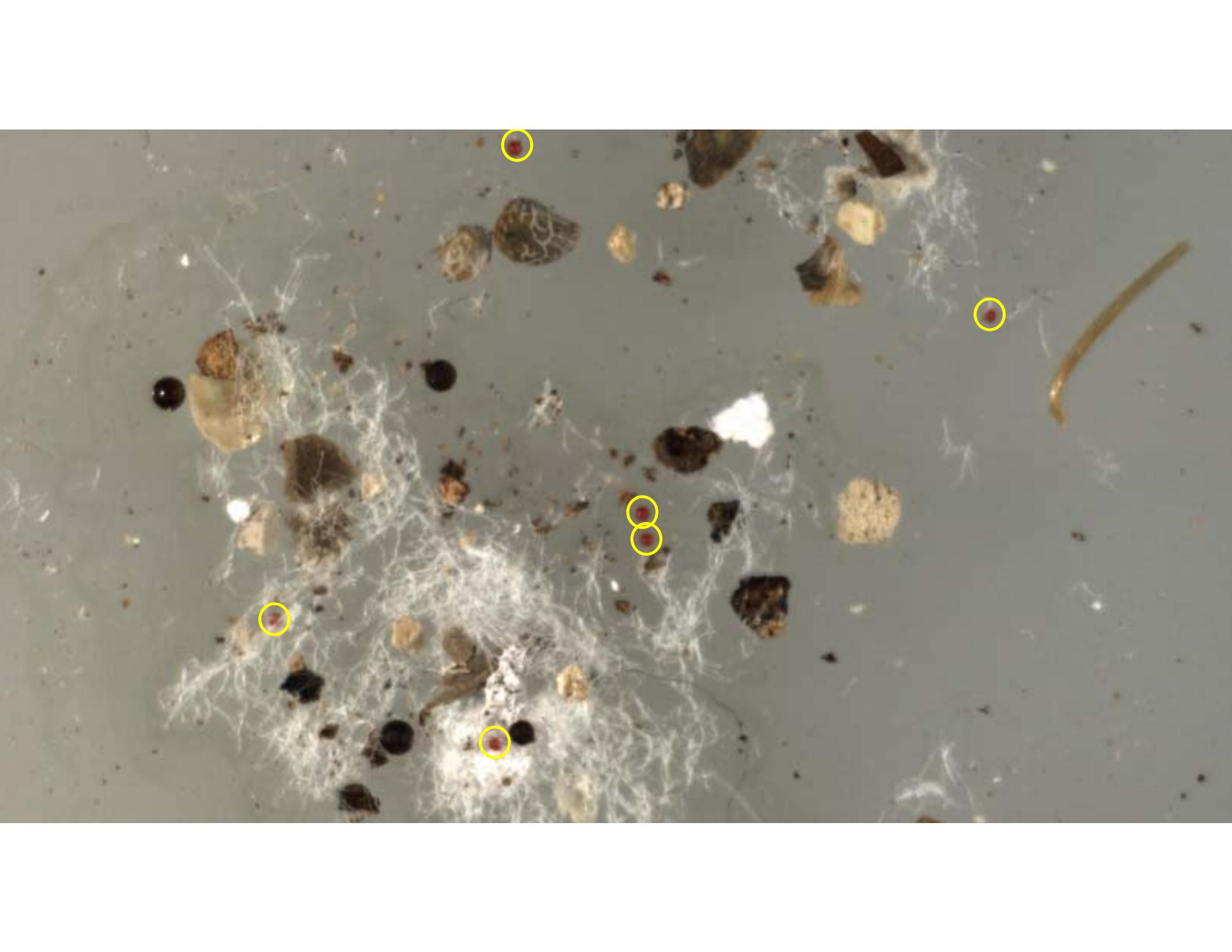
# Extraction Procedure

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1. Filter, dry & weigh sample **(24 hrs)**
2. Wet peroxide digestion **(24 hrs)**
3. Filter sample, protease digestion **(24 hrs)**
4. Filter sample, lipase digestion **(24 hrs)**
5. Filter sample, cellulase digestion **(24 hrs x 3)**
6. Filter sample, final wet peroxide, filter sample **(3 hrs)**





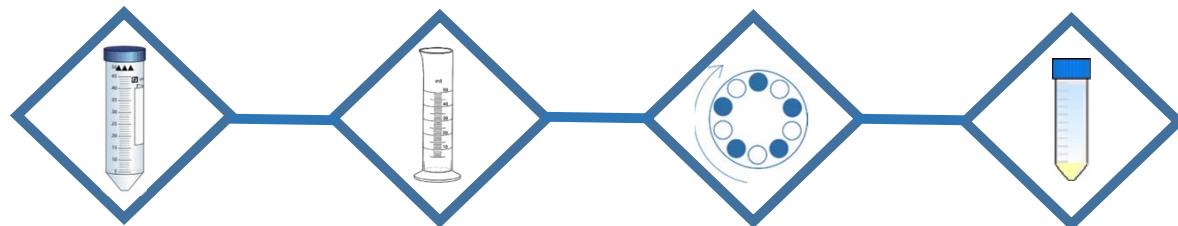


8

# Sample Preparation for Microplastics



Sample  
Preparation



- Easy to replicate
- Addresses Interferences
- Minimizes damage and loss of particles



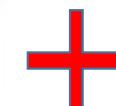
# Standardized Preparation

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Modified  
Schweizer's Reagent



Test Tube Shaker



Centrifuge



# Extraction Procedure

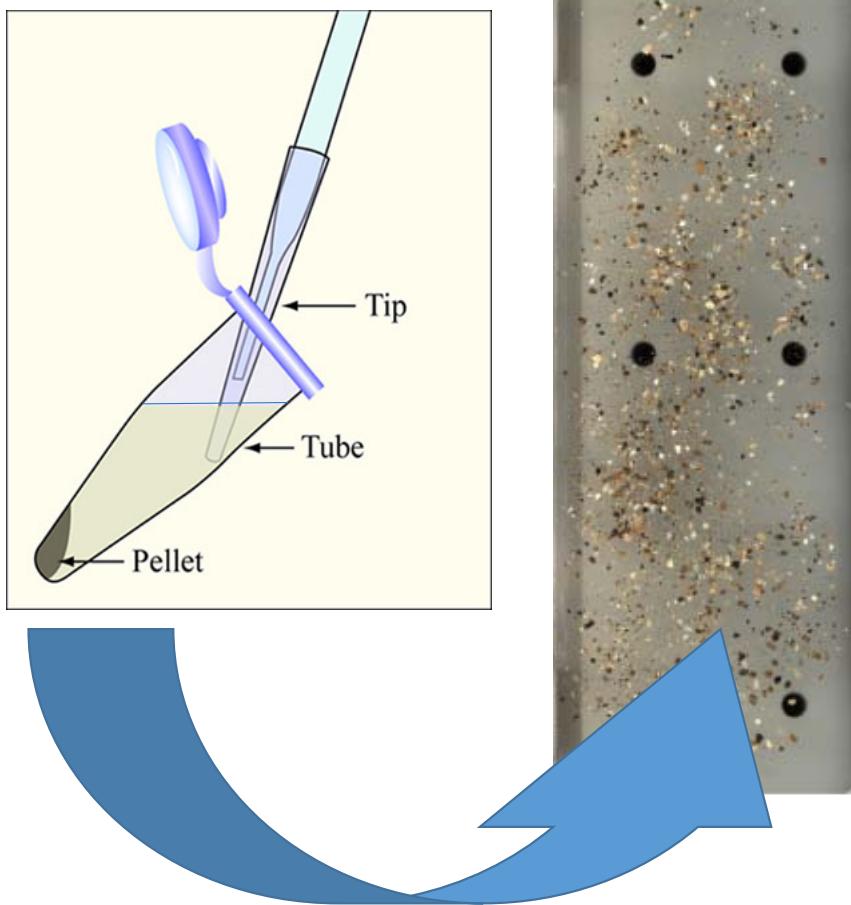
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1. Wet peroxide digestion (1 hr)
2. Centrifuge & decant excess liquid (15 min)
3. Cellulose digestion with modified Schweizer's Reagent (5 min)
4. Centrifuge & decant excess liquid (5 min)
5. Schweizer's Reagent quench with 30% NH<sub>4</sub>OH (5 min)
6. Buffer sample to pH 8 with Tris-HCl (5 min)
7. Simultaneous protease & lipase digestion (20 hrs)
8. Centrifuge & decant excess liquid (5min)
9. Water wash (5 min)
10. Centrifuge, & decant excess liquid, add 10 mL of CH<sub>4</sub>OH



## Observed Improvements

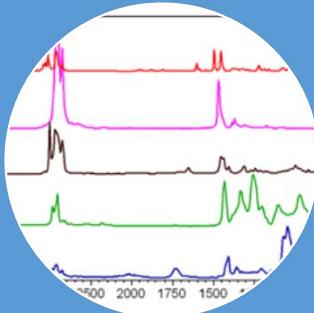
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- Eliminates filtration and associated sample loss
- Cellulose is completely removed during the first step
- Preparation time is ideal
- Entire sieved contents are plated onto a single slide
- Reference spike recovery near 100%

# Desired Characteristics

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Sample  
Analysis

1. Spectroscopic analysis
2. Identify targets
3. Isolate individual particles
4. Positive identification
5. Quantification of particle count and size

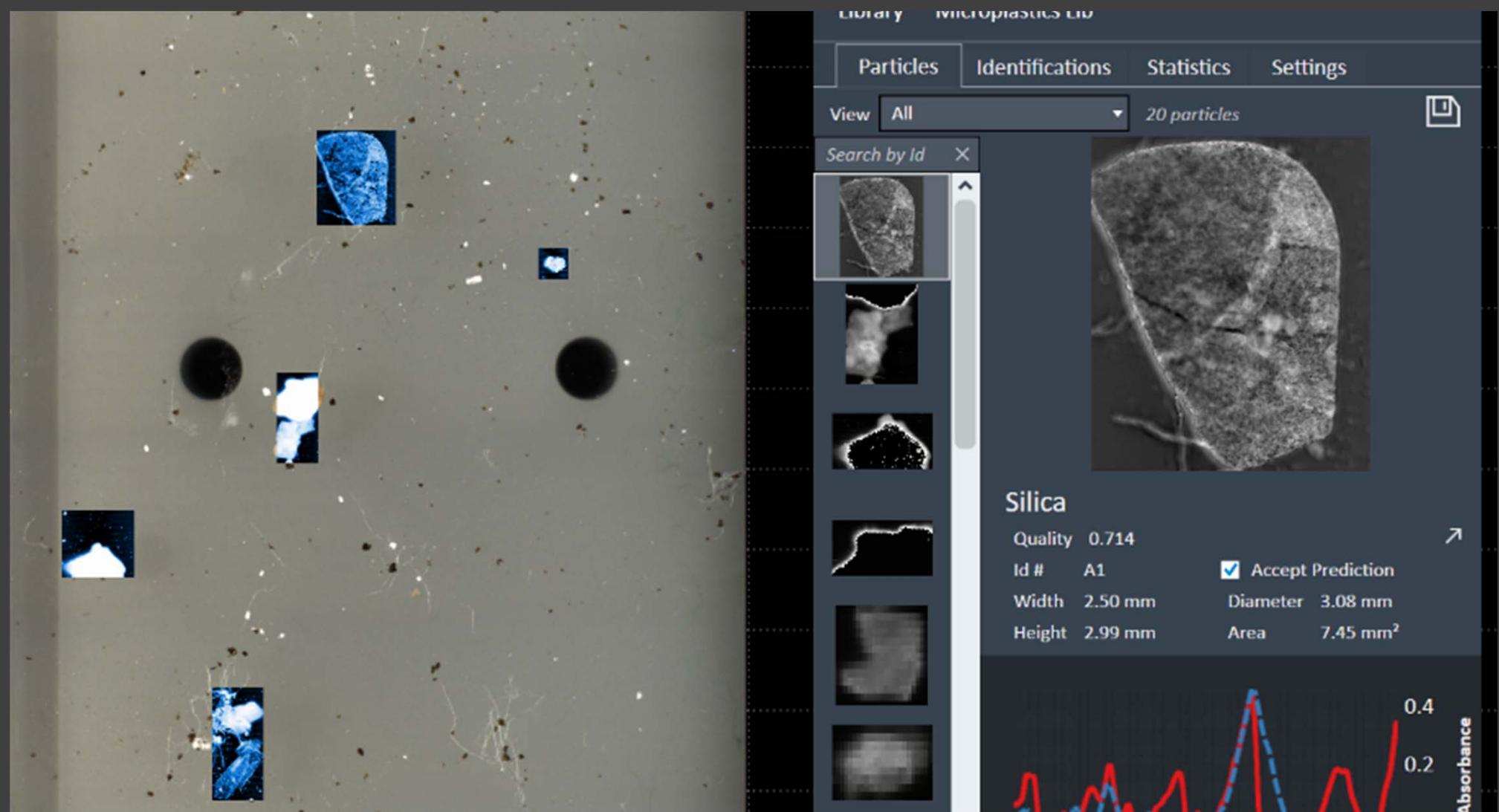


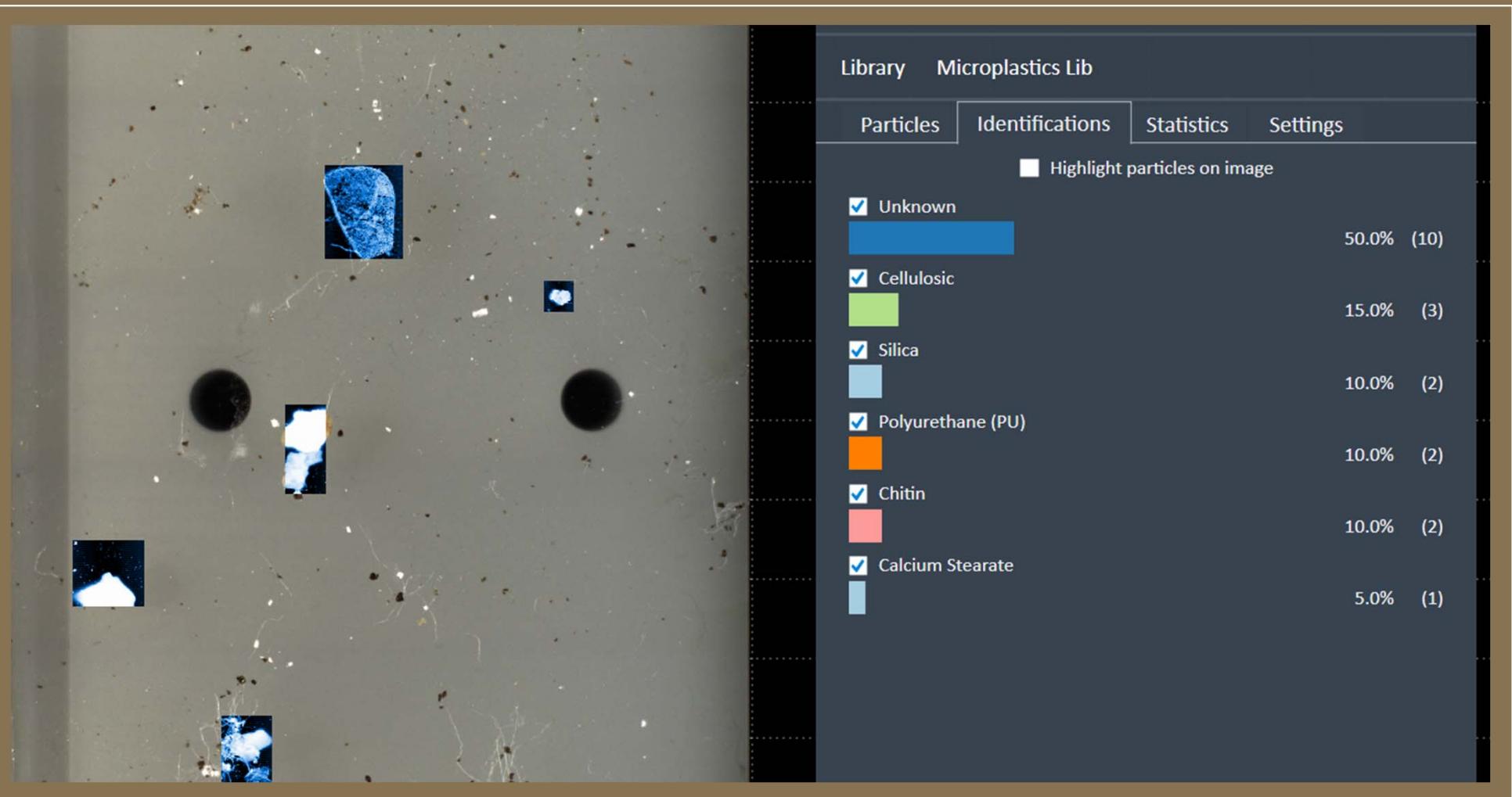
# Agilent LDIR 8700

## Chemical Imaging System

# Agilent LDIR 8700

Specification	Description
Optical Resolution	1 micron
Spectral Resolution	20 micron
Wavelength Coverage	1,800 – 1,000 wavenumbers (cm <sup>-1</sup> )
ATR Resolution	
Light Source	Quantum Cascade Laser
Detector	TE cooled MCT
Time Required for Analysis	Minutes
Reference Library	IR and buildable

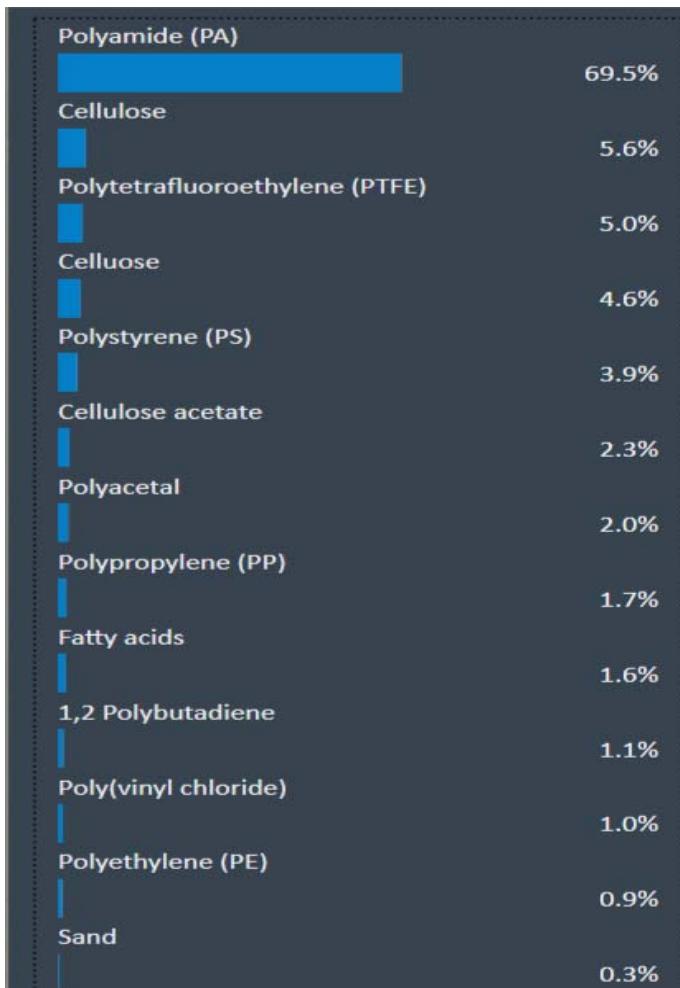




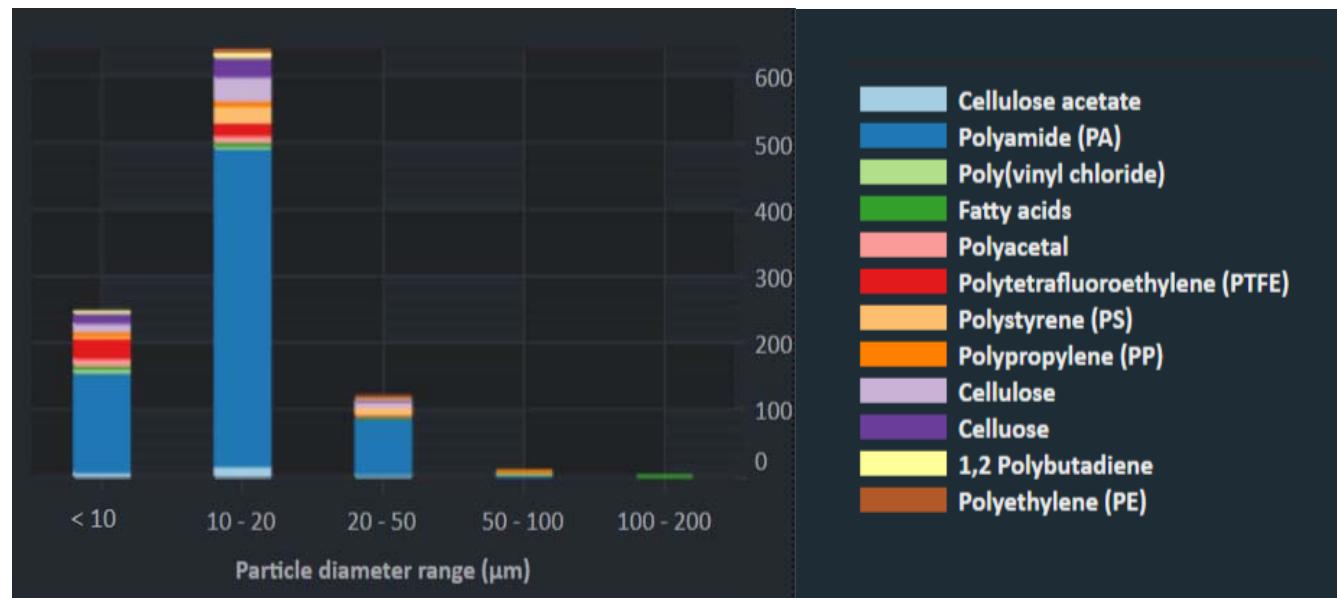
# Spectral Matching

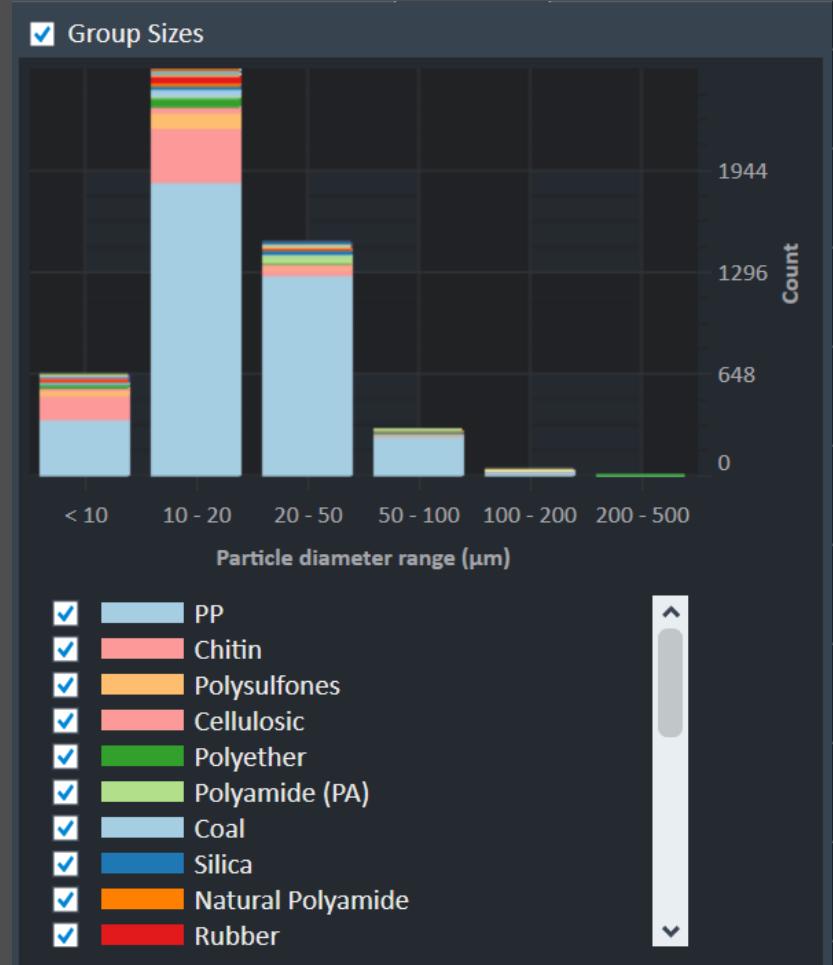
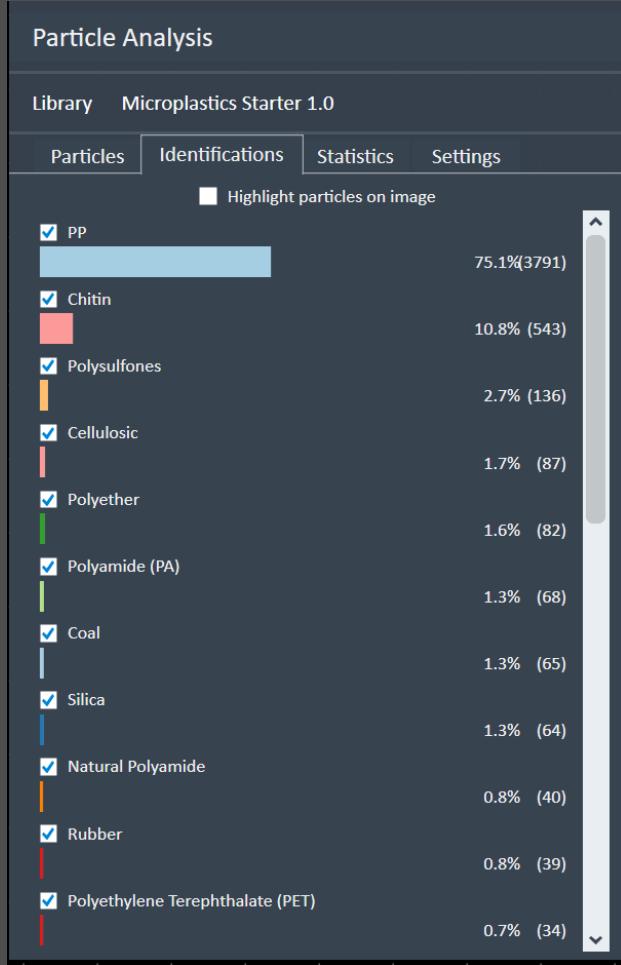


# Influent 300 micron



- 1028 particles detected
  - Polyamides were most prevalent
  - Wool, silk, nylon from textiles
  - Undigested protein-rich debris
  - Followed by cellulosic particles, PTFE (Teflon), polystyrene





# Instrument Validation

	<b>Sample Analyzed</b>	<b>LDIR Particle Count</b>	<b>Usable Particles Identified</b>	<b>Acrylonitrile Butadiene</b>	<b>Alkyd Varnish</b>	<b>Calcium Stearate</b>	<b>Cellulosic</b>	<b>Chitin</b>	<b>Coal</b>	<b>Natural Polyamide</b>	<b>Polyamide (PA)</b>	<b>Polyethylene (PE)</b>	<b>Polyethylene</b>	<b>Terephthalate (PET)</b>	<b>Polyimide</b>	<b>Polypropylene (PP)</b>	<b>Polytetrafluoroethylene (PTFE)</b>	<b>Polyurethane (PU)</b>	<b>Rubber</b>	<b>Silica</b>
	Run 1	136	132	7	5	4	12	1	0	60	9	2	2	1	0	9	1	19	1	0
	Run 2	138	131	3	1	5	12	1	1	63	7	2	1	0	9	1	25	0	0	
	Run 3	136	131	7	3	5	12	0	0	65	10	2	1	0	8	0	16	2	1	
	Run 4	140	135	7	6	5	12	0	1	63	11	2	1	1	8	1	17	0	0	
	Run 5	136	127	8	4	6	7	2	2	60	10	2	1	1	8	0	12	3	1	
	Run 6	137	130	1	1	5	12	1	2	60	11	2	2	0	8	0	24	1	0	
	Run 7	137	127	0	2	6	13	1	1	58	9	2	1	0	8	0	26	0	0	
	Run 8	135	135	0	3	6	11	1	3	58	9	1	0	1	9	1	25	1	0	
	True Number:														10					
	Mean Detected:	137	131	4.13	3.13	5.25	11.4	0.88	1.25	60.9	9.5	1.88	1.13	0.38	8.38	0.5	20.5	1	0.25	
	Percent Recovery:															83.8				
	Std Dev:	1.55	3.07	3.48	1.81	0.71	1.85	0.64	1.04	2.53	1.31	0.35	0.64	0.52	0.52	0.53	5.21	1.07	0.46	
	Relative Std Dev:	1.13	2.34	84.4	57.8	13.5	16.2	73.2	82.8	4.16	13.8	18.9	57	138	6.18	107	25.4	107	185	



## Particle Analysis

Library Microplastics Lib

Particles Identifications Statistics Settings

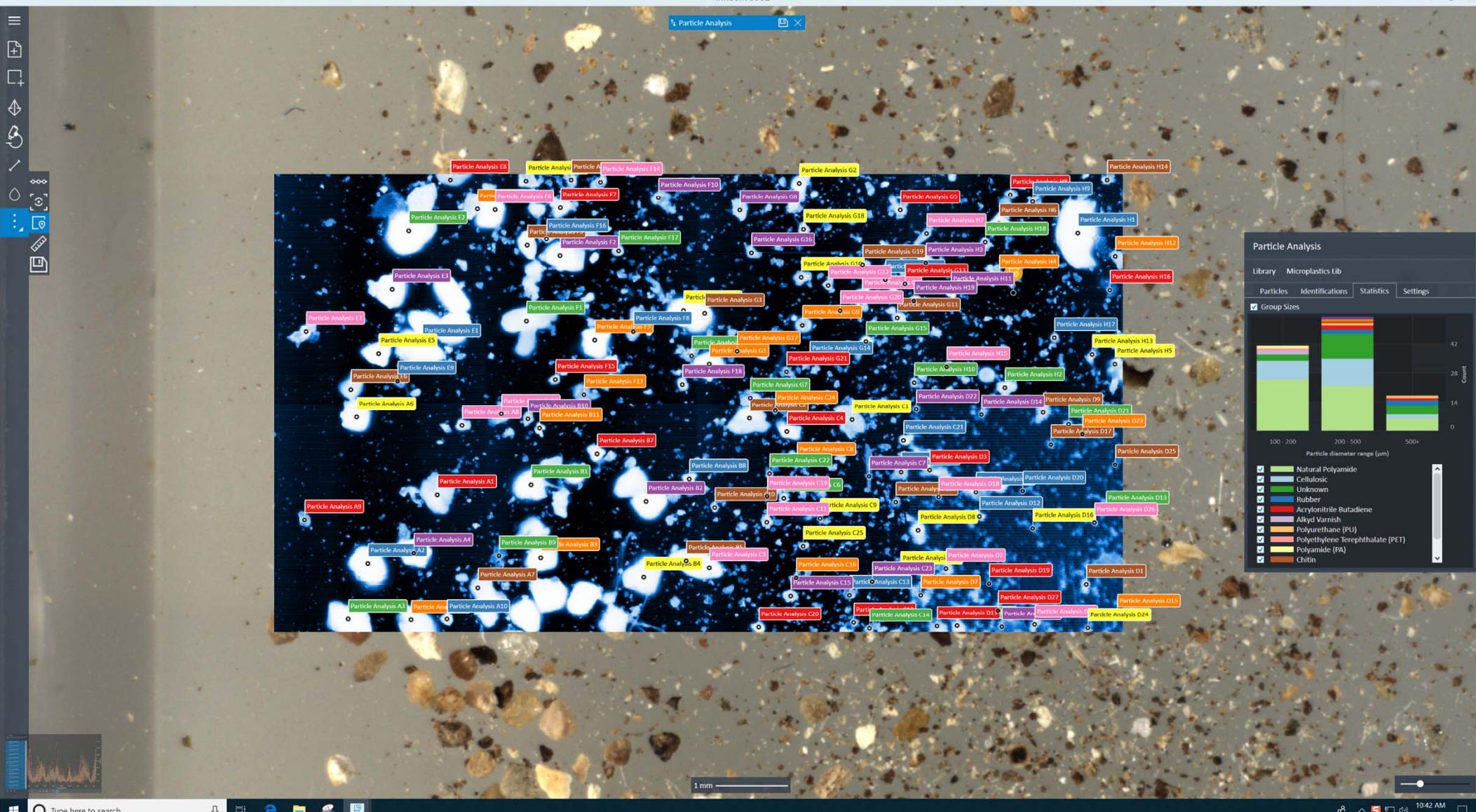
Auto Scan

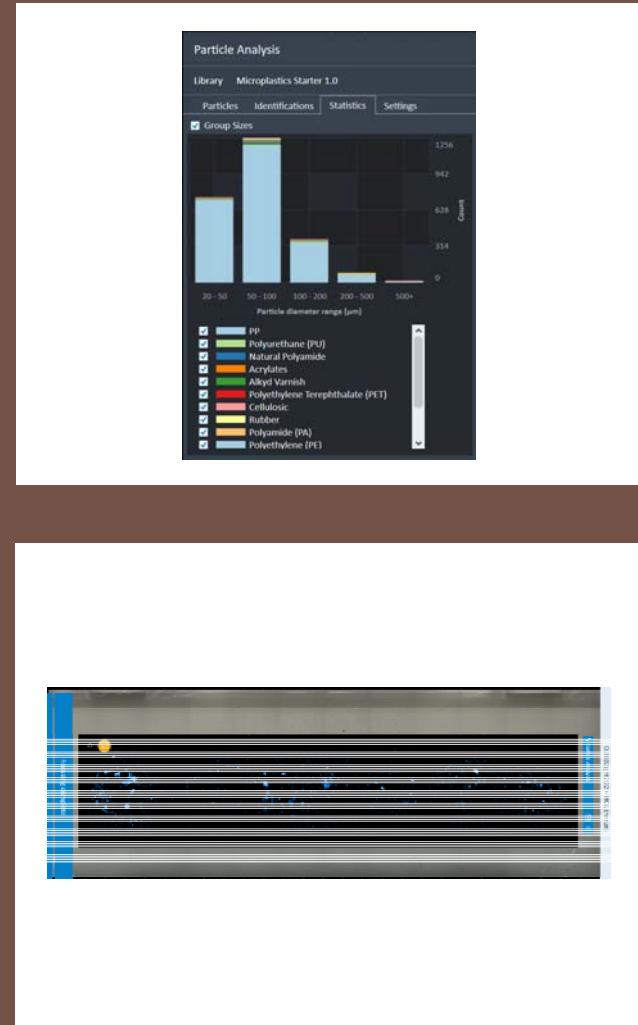
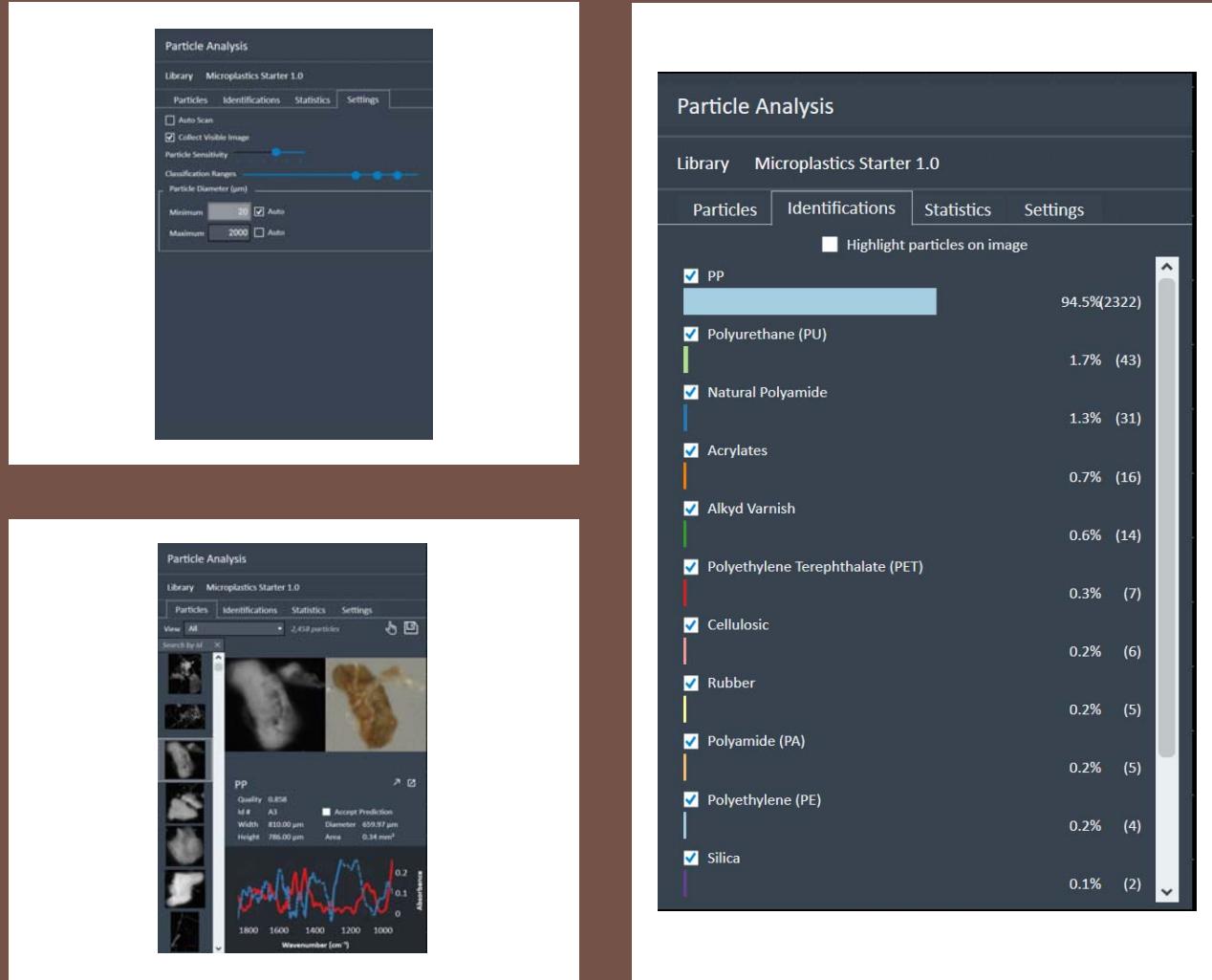
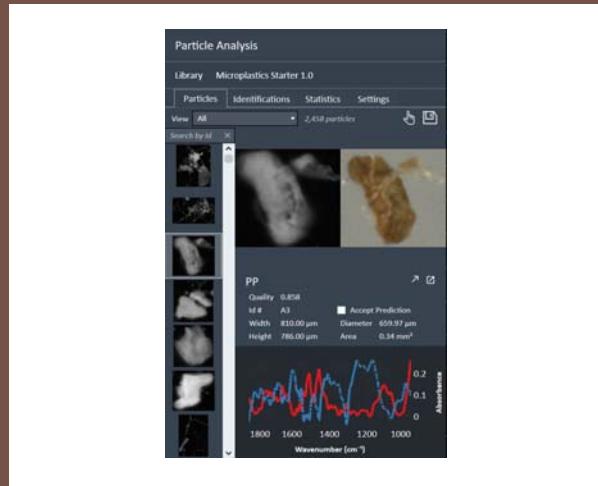
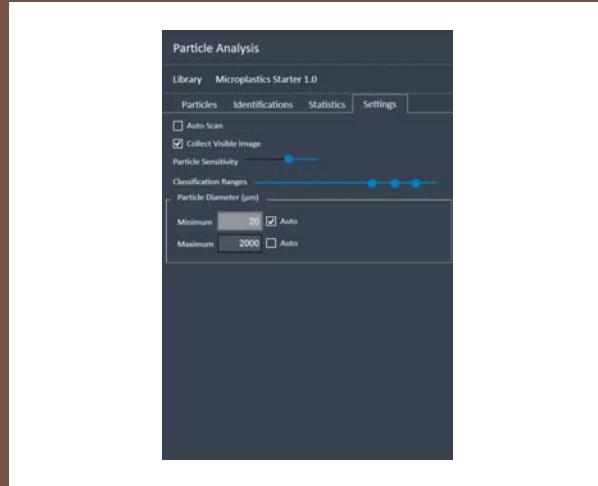
Particle Sensitivity

Particle Diameter ( $\mu\text{m}$ )

Minimum   Auto

Maximum   Auto



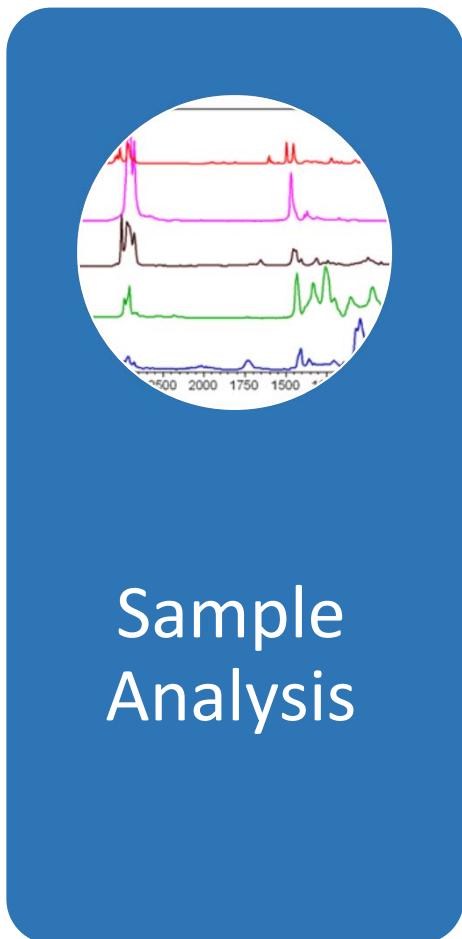


# Excel Data Export

#	Id	Width	Height	Diameter	Aspect Ratio	Area	Perimeter	Eccentricity	Circularity	Solidity	Identification	Quality	Is Valid
1	A1	721	529	540.5484	1.363601028	229487.5	2990.279552	0.794483	0.322512	0.780205	Cellulosic	0.70778	true
2	A2	589	494	535.8316	1.191709808	225500	1874.091612	0.664369	0.806817	0.969267	Rubber	0.686543	true
3	A3	625	430	524.1045	1.453488372	215737.5	1919.619399	0.617248	0.735708	0.970479	Natural Polyamide	0.699224	true
4	A4	657	415	473.4825	1.58367215	176075	1940.954526	0.581468	0.587323	0.896284	Unknown	0.626958	true
5	A5	550	375	443.7235	1.466666667	154637.5	1651.040754	0.570457	0.712868	0.974171	Unknown	0.644938	true
6	A6	430	395	425.4491	1.088607595	142162.5	1483.761536	0.696801	0.811459	0.967915	Cellulosic	0.669999	true
7	A7	230	805	393.5601	0.285714286	121650	2004.680365	0.863211	0.380392	0.880883	Unknown	0.630628	true
8	A8	210	167	186.5242	1.256410171	27325	648.7005711	0.671909	0.815984	0.96427	Natural Polyamide	0.69458	true
9	A9	216	154	177.8764	1.404301234	24850	622.8427076	0.544338	0.80497	0.965986	Polyethylene Terephthalate (PET)	0.653054	true
10	A10	182	169	174.7167	1.073770495	23975	599.4112474	0.720755	0.83853	0.974099	Natural Polyamide	0.699323	true
11	B1	897	816	750.3297	1.099287452	442175	4734.751766	0.725563	0.247862	0.750706	Natural Polyamide	0.682812	true
12	B2	906	629	683.9621	1.439613179	367412.5	3268.269461	0.526536	0.432243	0.756129	Unknown	0.647627	true
13	B3	770	492	591.9153	1.565899468	275175	2582.79218	0.545498	0.51837	0.850585	Acrylonitrile Butadiene	0.651467	true
14	B4	596	578	551.8541	1.031010139	239187.5	2116.015492	0.806203	0.671291	0.924262	Rubber	0.692184	true
15	B5	405	470	405.4714	0.861702128	129125	1608.11182	0.652729	0.627462	0.919285	Natural Polyamide	0.653315	true
16	B6	277	391	309.3025	0.708571501	75137.5	1132.045807	0.751547	0.736781	0.94468	Cellulosic	0.660203	true
17	B7	417	241	287.183	1.730526346	64775	1331.959578	0.584122	0.458812	0.857805	Natural Polyamide	0.670752	true
18	B8	262	220	211.3641	1.193002971	35087.5	967.9036713	0.790561	0.47065	0.755789	Natural Polyamide	0.666217	true
19	B9	305	167	177.1142	1.822222374	24637.5	1106.898619	0.729224	0.252692	0.689161	Natural Polyamide	0.74989	true
20	B10	159	208	170.7548	0.762520215	22900	642.8427058	0.792272	0.696363	0.906931	Natural Polyamide	0.708936	true
21	B11	155	172	155.8941	0.901204853	19087.5	612.3401785	0.756879	0.639696	0.879608	Natural Polyamide	0.663842	true
22	C1	840	500	554.4006	1.67999951	241400	3014.507912	0.664947	0.333821	0.813137	Natural Polyamide	0.718071	true
23	C2	825	435	527.8415	1.896551706	218825	2834.507915	0.865484	0.342256	0.777664	Natural Polyamide	0.656713	true
24	C3	600	446	447.0112	1.34631437	156937.5	2155.304814	0.713552	0.424541	0.759804	Natural Polyamide	0.683267	true
25	C4	363	336	310.0477	1.079202306	75500	1450.538229	0.700405	0.450919	0.80138	Natural Polyamide	0.684362	true
26	C5	255	403	293.2701	0.633367615	67550	1540.538228	0.810585	0.357676	0.798345	Cellulosic	0.771951	true
27	C6	269	588	283.4737	0.456880747	63112.5	1849.741321	0.811755	0.231795	0.605396	Natural Polyamide	0.725266	true
28	C7	149	667	253.8226	0.223171385	50600	1688.111818	0.958237	0.22313	0.647266	Natural Polyamide	0.708144	true
29	C8	422	173	237.2617	2.443395967	44212.5	1169.325024	0.564417	0.406335	0.833805	Polyurethane (PU)	0.712773	true

# LDIR Analysis of Microplastics

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Sample  
Analysis



- Identification of MP materials
- Quantification of MP particles
- Particle size distribution
- Reproducible

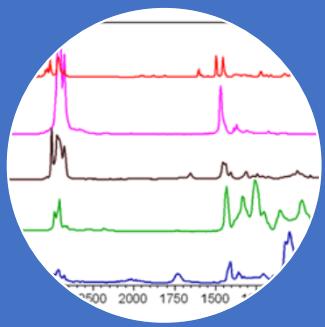




## Sample Collection



## Sample Preparation

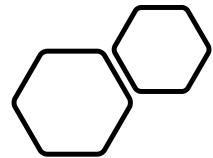


## Sample Analysis



Reproducibility Achieved





Thank You!

Stay Positive.....Test Negative

