Microbiology Expert Committee (MEC)
Meeting Summary

April 14, 2026

I Welcome and Roll Call:

Cody Danielson called the meeting to order at 1:32pm Eastern on April 14, 2026, by teleconference. Attendance is recorded in Attachment A – there were 8 members present. Associates present: Debbie Bond, Tiffany Carey, Thekkekalathil Chandrasekhar, Shannon Freedman, Hannah Herrick, Rachel Hook, Tammy Kreutzer, Morgan Lawrence, David Lo, Alma McCammond, Elisa Snyder, and Joshua Snyder. Paul Junio attended as Program Administrator.

II Approval of Agenda

Cody noted that the time for call-in information on the agenda was incorrect, and assured everyone that she isn't doing that intentionally (you had to be there – it's been an ongoing problem 😊). With that comment, the agenda was approved by consent.

III Discussion on Section 8.2.1.3 regarding types of water. Cody summarized the email traffic on this item and asked for input. Josh submitted alternate language for 8.2.1.3 b that was met with approval. Patsy suggested that 8.2.1.3 c should read 'in at-use volume'. Paul pointed out that the numbering is different in this draft (these sections are NOW 7.2.1.3) due to the removal of section 4. Robin noted that 7.2.1.2 e had similar language that needs the same time of changes as those made in the (new) 7.2.1.3.

7.2.1.3 Water types used in testing may include buffer water, peptone water, rinse water and/or reagent water.

a. For all laboratory-prepared water types used in testing that have a regulation-, method-, or standard-defined acceptable pH, the laboratory must verify and document the final pH prior to first use.

b. When types of water used in testing are purchased by the laboratory, the laboratory must review the certificate of analysis to verify that the pH is suitable for the method prior to first use.

c. When types of water used in testing are also purchased or prepared in at-use volume for dilutions, the laboratory must verify each working volume once per lot or prepared batch prior to first use.

Discussion on 7.2.1.2 – Robin attempted to capture the discussion regarding performance checks by separating positive and negative checks. A suggestion was made to say 'evidence of growth such as…' in 7.2.1.2 a. Comment was met with approval.

7.2.1.2 Performance Checks- All test reagents (as defined in 3.1) must be checked by the laboratory for satisfactory performance once per purchased lot or prepared batch prior to or in conjunction with first use.

a) Each test reagent must be analyzed with a positive control which verifies that the media/reagent produces the correct response. For selective media, the

positive must be a known which is pure (single organism/analyte of interest) and produce typical results. Non-selective media is used to determine that growth is possible and therefore evidence of growth is deemed a positive. As there is no specific target organism for non-selective media, any organism that produces evidence of growth, such as turbidity or colonies, would be deemed appropriate.

b) Each selective test reagent must be analyzed with a known negative control. A negative control demonstrates that the medium or test reagent does not support the growth of non-target organism(s)/analyte(s) of interest or does not exhibit the typical positive reaction of the target organism(s)/analyte(s) of interest.

c) When microorganisms are used for positive and negative controls for selective media and/or selective test reagents, the laboratory must use reference cultures traceable to national or international sources. . Microorganisms may be single-use preparations or cultures maintained for their intended use by demonstrated procedures that demonstrate the continued purity and viability of the organism.

i. Reference cultures, once prepared, may be sub-cultured once to provide reference stocks. The reference stocks must be preserved by a technique that maintains the characteristics of the strains. Working stocks must be prepared from reference stocks. If reference stocks have been thawed, they must not be refrozen and re-used.

ii. Working stocks must not be sequentially cultured more than five times. Each sequential culture must not be used beyond 31 days. Working stocks must not be sub-cultured to replace reference stocks.

d) To ensure accurate results, target organism identity must be verified as specified in the method (e.g., by use of the completed test, secondary verification tests such as a catalase test, or a selective medium such as Brilliant Green Lactose Bile Broth (BGLB) or EC or EC + MUG broth).

e) When prepared in the lab, the laboratory must verify and record the final pH of all media. For media purchased by the laboratory, the laboratory must review the certificate of analysis to verify that the pH is suitable for the method.

Response to comments document (these changes were captured in the document and are not produced here, but will be available in the document when posted):
Line 80 – persuasive - changing the order of 7.2.1.4 to edit/clarify
Line 81 – non-persuasive
Line 82 – persuasive, but not changed in the manner requested
Line 83 – persuasive comment per the Style Guide
Line 84 – non-persuasive
Line 85 – non-persuasive
Line 86 – persuasive – changed specification to requirement
Line 87 - persuasive – add ‘applicable’
Line 132 – non-persuasive – references to other Modules have been globally verified
Line 134 – persuasive – following discussion, the comment arrived at language regarding thermal preservation. This will no longer be covered by Module 2 as it makes sense for each Technical Module to address it specific to that module.

7.4.1 Samples which require thermal preservation but do not meet the maximum temperature requirement may be considered acceptable if appropriate evidence of sample cooling is present.

Line 137 – non-persuasive

Having reached the end of the scheduled time, Cody will send out a poll looking for a possible additional meeting time. *[EDITOR'S NOTE: the added meeting was scheduled for Monday, April 20 at 12PM Eastern. Meeting minutes from the March 10 meeting were distributed on April 16.]* Cody will send today's work products out and will summarize the Response to Comment document. The meeting adjourned at 3:06 PM Eastern.

Attachment A - Participants

Microbiology Expert Committee (MEC)

Name	Organization	Stakeholder	Email	Term Expires	Present
Nigel Allison (Vice Chair)	Los Angeles County Sanitation Districts	Lab	nallison@lacsd.org	2028*	X
Robin Cook	City of Daytona Beach	Lab	cookr@codb.us	2027	X
Cody Danielson (Chair)		Other	cody.danielson129@gmail.com	2028	X
Maria Fayard	ORELAP	AB	maria.j.fayard@oha.oregon.gov	2029	
Maria Friedman	California ELAP	AB	qamfriedman@gmail.com	2028	
Matt Graves	ERA	Other	matt_graves@waters.com	2028	
Silky Labie	ELCAT	Other	elcatllc@centurylink.net	2029	X
Liz Lesold	NYSDOH ELAP	AB	elizabeth.lesold@health.ny.gov	2027*	X
Brian Mercer	Brian Mercer	Lab	bmercer@plantation.org	2027*	X
Patsy Root	IDEXX Water	Other	patsy-root@idexx.com	2027*	X
Robert Royce	New Jersey DEP	AB	Robert.Royce@dep.nj.gov	2028	
Tina Shidel	Pace Analytical Services - Ormond Beach	Lab	tina.buttermore@pacelabs.com	2027*	X
Katie Strothman	Sanders Lab	Lab	katie@sanderslabs.net	2029	
Elizabeth Turner	Eurofins	Lab	elizabeth.turner@et.eurofinsus.com	2029	
Paul Junio	The NELAC Institute	NA	paul.junio@nelac-institute.org		X

* - eligible to serve another term